

Review

The histopathological and molecular heterogeneity of hepatocellular carcinoma: a narrative review

Wonju Chung, Haeryoung Kim

Department of Pathology, Seoul National University Hospital, Seoul National University
College of Medicine, Seoul, Korea

Running title: Heterogeneity of hepatocellular carcinoma

Corresponding author: Haeryoung Kim, Department of Pathology, Seoul National University
College of Medicine, 103 Daehak-no, Jongno-gu, Seoul, 03080, Korea, E-mail:
haeryoung.kim@snu.ac.kr

Abstract

Hepatocellular carcinoma (HCC) remains a leading cause of cancer-related deaths worldwide, with poor clinical outcomes due to challenges in early detection and limited efficacy of current treatments such as receptor tyrosine kinase inhibitors and immunotherapy. HCC exhibits significant heterogeneity at both histopathological and molecular levels, complicating its management but offering potential for personalized therapeutic approaches. This review outlines the morpho-molecular heterogeneity of HCC and summarizes various histological subtypes, including steatohepatic, clear cell, macrotrabecular-massive, scirrhous, lymphocyte-rich, and

fibrolamellar HCCs. Each subtype possesses distinct clinical, histological, and molecular features; for instance, steatohepatitic HCC is associated with metabolic dysfunction and shows IL-6/JAK/STAT activation, while clear cell HCCs often have IDH1 mutations and favorable prognosis. The macrotrabecular-massive subtype is linked to poor outcomes and TP53 mutations, whereas scirrhous HCCs express stemness markers and have TSC1/TSC2 mutations. Lymphocyte-rich HCCs are characterized by immune cell infiltration and better prognosis. CTNNB1-mutated HCCs show specific morphological features and may benefit from targeted therapies. Understanding these subtypes and associated molecular alterations is crucial for developing effective diagnostic and therapeutic strategies, including potential predictive biomarkers and personalized treatments. Additionally, the identification of patterns like vessels-encapsulating-tumor-clusters (VETC) offers prognostic implications and may guide therapeutic decisions. Recent molecular studies have enhanced our comprehension of HCC heterogeneity, laying the groundwork for more personalized approaches. Pathologists play a vital role in recognizing these subtypes, aiding in prognosis prediction and treatment planning. Advances in digital pathology and artificial intelligence may further facilitate biomarker research, ultimately improving patient outcomes in HCC management.

Keywords: Fatty liver; Hepatocellular carcinoma; Immunotherapy; Prognosis; Tyrosine kinase inhibitors

Introduction

Hepatocellular carcinoma (HCC) is the most common primary liver cancer, accounting for 75-80% of primary liver malignancies [1]. HCC mostly develops in the background of chronic liver disease, the most common etiologies being hepatitis B virus or hepatitis C virus (HCV) infection, and chronic alcohol abuse. During the recent year, metabolic syndrome has become another major risk factor for HCC, even in Asian countries [2,3].

The clinical outcome of patients with HCC is generally poor, mainly due to difficulties in early detection and limited treatment options for advanced disease [4,5]. Currently, systemic therapy, such as receptor tyrosine kinase inhibitors and immunotherapy, is the treatment of choice for the patients with unresectable HCC [5]. However, the efficacy of systemic therapy in HCC is still unsatisfactory with survival benefit of 1-3 months, objective response rate less than 30%, and a high incidence of adverse events [6,7]. In this context, understanding the heterogeneity of HCC will play a key role in developing effective diagnostic and therapeutic strategies, by offering potential predictive biomarkers and personalized approaches for HCC management.

Morpho-molecular heterogeneity of HCC

HCC is typically composed of tumor cells showing hepatocytic differentiation with variable degrees of atypia [8]. While HCC recapitulates the cytoarchitectural morphology of the liver to varying extents, there are alterations in the hepatic microarchitecture such as loss of portal tracts, reduction or loss of the reticulin framework, and thickening of the hepatic plates. Neoangiogenesis occurs in HCCs, in the form of sinusoidal capillarization and unpaired arteries [8]. These changes also result in the characteristic imaging features, such as the early enhancement on contrast enhanced computed tomography/magnetic resonance imaging (MRI).

However, there is considerable heterogeneity of HCC, in the microscopic growth patterns, cytological features, and histological grade. About 50% of HCCs demonstrate mixed patterns of growth: trabecular, pseudoglandular, solid, and macrotrabecular (Fig. 1) [9]. In addition, while most HCCs demonstrate cytological features that recapitulate those of normal hepatocytes (i.e. polygonal cells with abundant eosinophilic cytoplasm), some HCCs show extensive areas with clear cell change, fatty change, and cholestasis. Cytoplasmic inclusions (e.g., hyaline bodies, Mallory-Denk bodies, and pale bodies) may also be seen in some tumor cells [8]. Histological grading is currently performed according to either the four-tiered modified Edmondson and

Steiner system or the three-tiered World Health Organization (WHO) grading system [9-11].

Recent advances in genomic techniques have unraveled the heterogeneity in the mutational landscape of HCC [12,13]. The most frequently mutated genes include *TERT* promoter, *TP53*, *CTNNB1*, *ARID1A*, *ARID2*, *JAK1*, *ALB*, *AXIN1*, *NFE2L2*, and *RPS6KA3* [12]. In addition, gene expression profiling studies have suggested several molecular subclasses of HCC that correlate with the clinicopathological features, providing the foundation for an integrated morphological-molecular classification of HCC [14]. In the past two decades, there have been many efforts to establish a subclassification system that better categorizes HCCs with distinct clinical, histological, and molecular features (Table 1) [13,15-19]. HCC can be subclassified into two major groups, the proliferative class and the non-proliferative class. The proliferative class is characterized by high chromosomal instability and *TP53* mutations, and is associated with poor histological differentiation, frequent vascular invasion, increased alpha-fetoprotein (AFP) level, and overall poor clinical outcome [16,18]. On the other hand, the non-proliferative class displays chromosomal stability and a well-differentiated phenotype with less frequent vascular invasion [16,18]. *CTNNB1*-mutated HCCs belong to the latter group: these demonstrate frequent cholestasis and less immune cell infiltration on histology [14,20].

Currently, approximately 35% of HCC can be further subclassified into histological subtypes with distinct morphological, clinicopathological and molecular characteristics (Table 1) [9]. The following section will summarize the clinicopathological and molecular features of these different subtypes.

Steatohepatic HCC

The steatohepatic subtype of HCC demonstrates the key histological features of non-neoplastic steatohepatitis, including steatosis, pericellular fibrosis, cell ballooning, inflammation, and Mallory-Denk bodies, and these features occupy a major portion (>50%) of the tumor (Fig. 2) [21].

This subtype has been more frequently identified in patients with underlying metabolic dysfunction-associated steatotic liver disease and alcohol abuse, and its relative frequency has been reported to be between 5 and 20% [3,21]. Steatohepatic HCC has been associated with less frequent vascular invasion and satellite nodules; however, its prognosis appears to be similar to that of conventional HCC [14]. Key molecular alterations associated with steatohepatic HCC include IL-6/JAK/STAT activation, while *CTNNB1*, *TERT* promoter and *TP53* mutations have been found to be less frequent in these tumors [14].

Clear cell HCC

By definition, in clear cell HCCs, more than 80% of tumor cells demonstrate abundant clear cytoplasm (Fig. 3). The clear cytoplasm is a result of glycogen accumulation; however, some tumor cells may appear clear due to lipid droplets, and some degree of steatosis is acceptable for this diagnosis [22]. The relative frequency of clear cell HCC has been estimated to be around 3-7%. Clear cell HCCs are generally well-differentiated tumors with a favorable prognosis [23]. One study has reported that clear cell HCCs demonstrate higher frequency of *IDH1* mutation, although this mutation is not sufficient to define the subtype [24].

Macrotrabecular-massive HCC

The macrotrabecular-massive subtype of HCC is an HCC in which more than 50% of the tumor cells assume a macrotrabecular growth pattern, defined as large trabeculae that are more than 6-10 cells thick (Fig. 4) [25]. This subtype accounts for approximately 5% of all HCCs and has been strongly associated with elevated serum AFP levels, high-grade cytological atypia, extensive lymphovascular invasion, more frequent distant metastasis, and a poor prognosis [25,26]. In addition, the vessels-encapsulating-tumor-clusters (VETC) pattern of neoangiogenesis, which has been associated with metastatic dissemination of HCC, is often enriched in this subtype (Fig.

4) [27,28]. *TP53* mutations and *FGF19* amplifications have been more frequently identified in the macrotrabecular-massive subtype of HCC [14].

Scirrhous HCC

This subtype is characterized by dense intratumoral fibrous stroma (Fig. 5). The scirrhous subtype has a relative frequency of 4% and often mimics intrahepatic cholangiocarcinoma on imaging [29]. Expression of immunohistochemical markers associated with stemness (e.g. cytokeratin (CK) 7, CK19, and epithelial cell adhesion molecule) is often seen in scirrhous HCCs, and increased expression of cholangiocarcinoma-like and stem-cell-like genes have been identified by gene expression profiling, consistent with the intermediate characteristic of this subtype [30,31]. Furthermore, scirrhous HCC is associated with frequent *TSC1/TSC2* mutations and transforming growth factor- β signaling activation [14,30].

Lymphocyte-rich HCC

The lymphocyte-rich subtype demonstrates massive intratumoral infiltration of lymphocytes, which outnumber the tumor cells in most microscopic fields. This subtype is rare, accounting for less than 1% of all HCCs, but has received much attention as it has been associated with a favorable clinical outcome [32]. The lymphocyte-rich subtype is associated with increased programmed death-ligand 1 expression and focal amplification of chromosome 11q13.3, which is related to the immune checkpoint signature (*CD274*, *PDCD1*, *BTLA*, *CTLA4*, *HAVCR2*, *IDO1*, and *LAG3*) [32-35]. Interestingly, although this subtype is also known as “lymphoepithelioma-like HCC”, it is not associated with Epstein-Barr virus infection, unlike the lymphoepithelioma-like tumors arising in other organs, such as the nasopharynx and stomach [32].

Fibrolamellar HCC

Fibrolamellar carcinoma, or fibrolamellar HCC, consists of strands of large eosinophilic tumor cells with abundant cytoplasm and prominent nucleoli, and separated by dense intratumoral bands of fibrosis [36]. Fibrolamellar carcinoma accounts for approximately 1% of all HCC, occurs in younger patients (median age of 25 years), and the background liver is non-cirrhotic [37]. The prognosis of fibrolamellar carcinoma appears to be better than that of conventional HCC arising in cirrhotic livers, but similar to that of HCC in non-cirrhotic livers [37]. *DNAJB1-PRKACA* gene fusion has been identified in >95% of cases, and fluorescence in situ hybridization (FISH) for *PRKACA* gene rearrangement is a useful ancillary test in confirming the diagnosis [38]. Expression of CK7 and CD68 in the tumor cells is another characteristic of fibrolamellar carcinoma [38].

***CTNNB1*-mutated HCC**

CTNNB1 mutations have been reported in approximately 20-40% of HCCs [39]. *CTNNB1* encodes β -catenin, which plays a key role in the WNT signaling pathway that regulates liver function and zonation [40]. In addition, bile salt transporter expression is dysregulated in these tumors, histologically manifested by frequent intratumoral cholestasis. Some *CTNNB1*-mutated HCCs may be diagnosed by gadoxetic acid-enhanced MRI, due to the upregulation of the organic anion transporting polypeptide 1B3 (OATP1B3) [41]. Histologically, *CTNNB1*-mutated HCCs are typically well-differentiated tumors with microtrabecular and/or pseudoglandular growth patterns, intratumoral cholestasis, and less immune cell infiltration compared to non-*CTNNB1*-mutated HCCs [14,20]. However, *CTNNB1*-mutated HCCs are not morphologically homogeneous, with approximately 40% not demonstrating the “classic *CTNNB1* morphology” [42]. Immunohistochemical studies for β -catenin (nuclear expression) and glutamine synthetase (diffuse, strong and homogeneous expression) may serve as useful surrogate

markers for *CTNNB1* mutation.

HCC with stemness-related marker expression

HCC with stemness-related marker expression, or progenitor HCC, is defined as HCC expressing stemness-related markers, e.g. CK19, in >5% of the tumor cells [15]. This subset of HCCs differ from combined hepatocellular-cholangiocarcinoma, as they are morphologically compatible with HCC, and there is no evidence of glandular differentiation or mucin production in these tumors. They are associated with increased serum AFP levels, frequent vascular invasion, poor histological differentiation, high recurrence rate, resistance to systemic chemotherapy and locoregional treatment, and overall poor prognosis [43]. HCCs with stemness-related marker expression more frequently demonstrate TP53 mutations and chromosomal instability, and increased PD-L1 expression [33,34].

HCCs with vessels-encapsulating-tumor-clusters (VETC) pattern

The VETC phenotype is defined by the presence of VETC pattern in more than 55% of the tumor area, characterized by CD34-positive vessels that encapsulate and isolate individual tumor clusters, forming a cobweb-like pattern (Fig. 4) [27,28,44]. The VETC pattern is often found in the macrotrabecular-massive subtype of HCC (7.8%) and is associated with aggressive behavior and metastatic dissemination [27,28]. It has been reported that VETC pattern is related to a novel mechanism of metastasis, independent of epithelial-to-mesenchymal transition [44]. Furthermore, patients with VETC-positive HCC have shown greater survival benefits from sorafenib therapy compared to those with VETC-negative HCC, suggesting that the VETC pattern may serve as a potential predictive marker for sorafenib response [28]. Correlation between the VETC pattern on histology and a rim arterial phase hyperenhancement in arterial phase imaging suggests a role for imaging in the prognostication of HCC [45].

Other rare histological subtypes of HCC have been described. The chromophobe subtype of HCC has tumor cells with clear to pale cytoplasm and mainly bland nuclei with focal areas of striking nuclear atypia [46]. Chromophobe subtype is strongly associated with alternative lengthening of telomeres, a telomerase-independent mechanism of telomere maintenance, which can be detected by fluorescence in situ hybridization [46]. Its prognosis is currently known to be similar to that of conventional HCC [47]. Neutrophil-rich HCC is characterized by marked intratumoral neutrophilic infiltration, granulocyte colony-stimulating factor production by tumor cells, and a poor prognosis [47]. The tumor cells are often poorly differentiated, and focal sarcomatoid differentiation can be observed [48].

Conclusion

Recent molecular studies have significantly enhanced our understanding of the morphological and molecular heterogeneity of HCC, providing the foundation for more effective and personalized treatment strategies. Pathologists are becoming increasingly aware of the histomorphological heterogeneity of HCC, and the specification of the various subtypes of HCC has helped pathologists understand the histology of HCC in more detail and the various differential diagnoses and diagnostic pitfalls for each variant. The correlation between the histomorphology and the molecular and biological features suggests the role of histology in the prediction of therapeutic response and prognosis. This may be further facilitated by the recent advances in digital pathology and artificial intelligence-based biomarker research.

ORCID

Wonju Chung: <https://orcid.org/0009-0006-8239-9968>

Haeryoung Kim: <https://orcid.org/0000-0002-4205-9081>

Authors' contributions

Conceptualization: Chung W, Kim H

Original draft preparation: Chung W,

Review and editing: Chung W, Kim H

Conflicts of interest

No potential conflict of interest relevant to this article was reported. Ethics statement: not applicable.

Funding

This was supported by the National Research Foundation of Korea (NRF) grant funded by the Korea government (MSIT) (NRF-2022R1A2C2010348).

Data availability

Not applicable.

Acknowledgments

Not applicable.

Supplementary materials

Not applicable.

References

1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA Cancer J Clin* 2018;68(6):394-424.
2. Kim DY. Changing etiology and epidemiology of hepatocellular carcinoma: Asia and worldwide. *J Liver Cancer* 2024;24(1):62-70.
3. Daher D, Dahan KSE, Singal AG. Non-alcoholic fatty liver disease-related hepatocellular carcinoma. *J Liver Cancer* 2023;23(1):127-142.
4. Forner A, Llovet JM, Bruix J. Hepatocellular carcinoma. *Lancet* 2012;379(9822):1245-1255.
5. Gordan JD, Kennedy EB, Abou-Alfa GK, Beg MS, Brower ST, Gade TP, et al. Systemic Therapy for Advanced Hepatocellular Carcinoma: ASCO Guideline. *J Clin Oncol* 2020;38(36):4317-4345.
6. Doycheva I, Thuluvath PJ. Systemic Therapy for Advanced Hepatocellular Carcinoma: An Update of a Rapidly Evolving Field. *J Clin Exp Hepatol* 2019;9(5):588-596.
7. Song YG, Yoo JJ, Kim SG, Kim YS. Complications of immunotherapy in advanced hepatocellular carcinoma. *J Liver Cancer* 2024;24(1):9-16.
8. Burt AD, Ferrell LD, Hübscher SG. MacSween's Pathology of the Liver Elsevier Health Sciences; 2022.
9. Nagtegaal ID, Odze RD, Klimstra D, Paradis V, Rugge M, Schirmacher P, et al. The 2019 WHO classification of tumours of the digestive system. *Histopathology* 2020;76(2):182-188.
10. Edmondson HA, Steiner PE. Primary carcinoma of the liver: a study of 100 cases among 48,900 necropsies. *Cancer* 1954;7(3):462-503.
11. Martins-Filho SN, Paiva C, Azevedo RS, Alves VAF. Histological Grading of Hepatocellular Carcinoma-A Systematic Review of Literature. *Front Med (Lausanne)* 2017;4:193.
12. Lee JS. The mutational landscape of hepatocellular carcinoma. *Clin Mol Hepatol* 2015;21(3):220-229.
13. Cancer Genome Atlas Research Network. Electronic address wbe, Cancer Genome Atlas Research N. Comprehensive and Integrative Genomic Characterization of Hepatocellular Carcinoma. *Cell* 2017;169(7):1327-1341 e1323.
14. Calderaro J, Couchy G, Imbeaud S, Amaddeo G, Letouze E, Blanc JF, et al. Histological subtypes of hepatocellular carcinoma are related to gene mutations and molecular tumour classification. *J Hepatol* 2017;67(4):727-738.
15. Lee JS, Heo J, Libbrecht L, Chu IS, Kaposi-Novak P, Calvisi DF, et al. A novel prognostic subtype of human hepatocellular carcinoma derived from hepatic progenitor cells. *Nat*

- Med* 2006;12(4):410-416.
16. Boyault S, Rickman DS, de Reyniès A, Balabaud C, Rebouissou S, Jeannot E, et al. Transcriptome classification of HCC is related to gene alterations and to new therapeutic targets. *Hepatology* 2007;45(1):42-52.
 17. Chiang DY, Villanueva A, Hoshida Y, Peix J, Newell P, Minguez B, et al. Focal gains of VEGFA and molecular classification of hepatocellular carcinoma. *Cancer Res* 2008;68(16):6779-6788.
 18. Hoshida Y, Nijman SM, Kobayashi M, Chan JA, Brunet JP, Chiang DY, et al. Integrative transcriptome analysis reveals common molecular subclasses of human hepatocellular carcinoma. *Cancer Res* 2009;69(18):7385-7392.
 19. Sia D, Jiao Y, Martinez-Quetglas I, Kuchuk O, Villacorta-Martin C, Castro de Moura M, et al. Identification of an Immune-specific Class of Hepatocellular Carcinoma, Based on Molecular Features. *Gastroenterology* 2017;153(3):812-826.
 20. Audard V, Grimber G, Elie C, Radenen B, Audebourg A, Letourneur F, et al. Cholestasis is a marker for hepatocellular carcinomas displaying β -catenin mutations. *The Journal of Pathology* 2007;212(3):345-352.
 21. Salomao M, Yu WM, Brown RSJ, Emond JC, Lefkowitz JH. Steatohepatic Hepatocellular Carcinoma (SH-HCC): A Distinctive Histological Variant of HCC in Hepatitis C Virus-related Cirrhosis With Associated NAFLD/NASH. *The American Journal of Surgical Pathology* 2010;34(11):1630-1636.
 22. Bannasch P, Ribback S, Su Q, Mayer D. Clear cell hepatocellular carcinoma: origin, metabolic traits and fate of glycogenotic clear and ground glass cells. *Hepatobiliary & Pancreatic Diseases International* 2017;16(6):570-594.
 23. Li T, Fan J, Qin L-X, Zhou J, Sun H-C, Qiu S-J, et al. Risk Factors, Prognosis, and Management of Early and Late Intrahepatic Recurrence After Resection of Primary Clear Cell Carcinoma of the Liver. *Annals of Surgical Oncology* 2011;18(7):1955-1963.
 24. Lee JH, Shin DH, Park WY, Shin N, Kim A, Lee HJ, et al. IDH1 R132C mutation is detected in clear cell hepatocellular carcinoma by pyrosequencing. *World J Surg Oncol* 2017;15(1):82.
 25. Jeon Y, Benedict M, Taddei T, Jain D, Zhang X. Macrotrabecular Hepatocellular Carcinoma: An Aggressive Subtype of Hepatocellular Carcinoma. *Am J Surg Pathol* 2019;43(7):943-948.
 26. Kumar D, Hafez O, Jain D, Zhang X. Can primary hepatocellular carcinoma histomorphology predict extrahepatic metastasis? *Hum Pathol* 2021;113:39-46.
 27. Renne SL, Woo HY, Allegra S, Rudini N, Yano H, Donadon M, et al. Vessels Encapsulating Tumor Clusters (VETC) Is a Powerful Predictor of Aggressive Hepatocellular Carcinoma. *Hepatology* 2020;71(1):183-195.
 28. Fang JH, Xu L, Shang LR, Pan CZ, Ding J, Tang YQ, et al. Vessels That Encapsulate Tumor

- Clusters (VETC) Pattern Is a Predictor of Sorafenib Benefit in Patients with Hepatocellular Carcinoma. *Hepatology* 2019;70(3):824-839.
29. Kurogi M, Nakashima O, Miyaaki H, Fujimoto M, Kojiro M. Clinicopathological study of scirrhous hepatocellular carcinoma. *J Gastroenterol Hepatol* 2006;21(9):1470-1477.
 30. Seok JY, Na DC, Woo HG, Roncalli M, Kwon SM, Yoo JE, et al. A fibrous stromal component in hepatocellular carcinoma reveals a cholangiocarcinoma-like gene expression trait and epithelial-mesenchymal transition. *Hepatology* 2012;55(6):1776-1786.
 31. Matsuura S, Aishima S, Taguchi K, Asayama Y, Terashi T, Honda H, et al. 'Scirrhous' type hepatocellular carcinomas: a special reference to expression of cytokeratin 7 and hepatocyte paraffin 1. *Histopathology* 2005;47(4):382-390.
 32. Chan AW, Tong JH, Pan Y, Chan SL, Wong GL, Wong VW, et al. Lymphoepithelioma-like hepatocellular carcinoma: an uncommon variant of hepatocellular carcinoma with favorable outcome. *Am J Surg Pathol* 2015;39(3):304-312.
 33. Calderaro J, Rousseau B, Amaddeo G, Mercey M, Charpy C, Costentin C, et al. Programmed death ligand 1 expression in hepatocellular carcinoma: Relationship With clinical and pathological features. *Hepatology* 2016;64(6):2038-2046.
 34. Nishida N, Sakai K, Morita M, Aoki T, Takita M, Hagiwara S, et al. Association between Genetic and Immunological Background of Hepatocellular Carcinoma and Expression of Programmed Cell Death-1. *Liver Cancer* 2020;9(4):426-439.
 35. Chan AW, Zhang Z, Chong CC, Tin EK, Chow C, Wong N. Genomic landscape of lymphoepithelioma-like hepatocellular carcinoma. *J Pathol* 2019;249(2):166-172.
 36. Torbenson M. Fibrolamellar carcinoma: 2012 update. *Scientifica (Cairo)* 2012;2012:743790.
 37. El-Serag HB, Davila JA. Is fibrolamellar carcinoma different from hepatocellular carcinoma? A US population-based study. *Hepatology* 2004;39(3):798-803.
 38. Graham RP, Yeh MM, Lam-Himlin D, Roberts LR, Terracciano L, Cruise MW, et al. Molecular testing for the clinical diagnosis of fibrolamellar carcinoma. *Mod Pathol* 2018;31(1):141-149.
 39. Gougelet A, Torre C, Veber P, Sartor C, Bachelot L, Denechaud PD, et al. T-cell factor 4 and beta-catenin chromatin occupancies pattern zonal liver metabolism in mice. *Hepatology* 2014;59(6):2344-2357.
 40. Monga SP. β -Catenin Signaling and Roles in Liver Homeostasis, Injury, and Tumorigenesis. *Gastroenterology* 2015;148(7):1294-1310.
 41. Ueno A, Masugi Y, Yamazaki K, Komuta M, Effendi K, Tanami Y, et al. OATP1B3 expression is strongly associated with Wnt/ β -catenin signalling and represents the transporter of gadoxetic acid in hepatocellular carcinoma. *J Hepatol* 2014;61(5):1080-1087.

42. Torbenson M, McCabe CE, O'Brien DR, Yin J, Bainter T, Tran NH, et al. Morphological heterogeneity in beta-catenin-mutated hepatocellular carcinomas: implications for tumor molecular classification. *Human Pathology* 2022;119:15-27.
43. Rhee H, Kim H, Park YN. Clinico-Radio-Pathological and Molecular Features of Hepatocellular Carcinomas with Keratin 19 Expression. *Liver Cancer* 2020;9(6):663-681.
44. Fang JH, Zhou HC, Zhang C, Shang LR, Zhang L, Xu J, et al. A novel vascular pattern promotes metastasis of hepatocellular carcinoma in an epithelial-mesenchymal transition-independent manner. *Hepatology* 2015;62(2):452-465.
45. Hwang SH, Rhee H. Radiologic features of hepatocellular carcinoma related to prognosis. *J Liver Cancer* 2023;23(1):143-156.
46. Wood LD, Heaphy CM, Daniel HD, Naini BV, Lassman CR, Arroyo MR, et al. Chromophobe hepatocellular carcinoma with abrupt anaplasia: a proposal for a new subtype of hepatocellular carcinoma with unique morphological and molecular features. *Mod Pathol* 2013;26(12):1586-1593.
47. Torbenson MS. Hepatocellular carcinoma: making sense of morphological heterogeneity, growth patterns, and subtypes. *Hum Pathol* 2021;112:86-101.
48. Amano H, Itamoto T, Emoto K, Hino H, Asahara T, Shimamoto F. Granulocyte colony-stimulating factor-producing combined hepatocellular/cholangiocellular carcinoma with sarcomatous change. *J Gastroenterol* 2005;40(12):1158-1159.

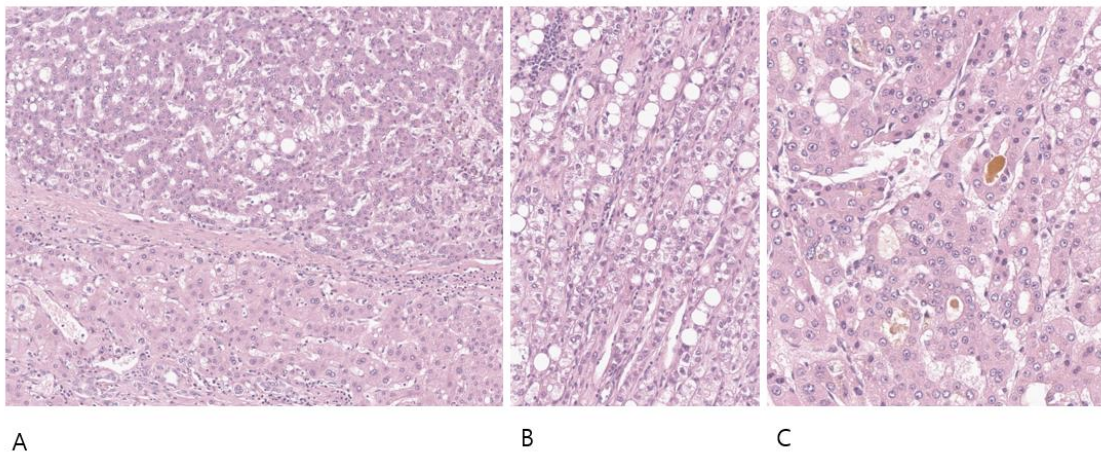
Table 1. Integrated morphological-molecular classification of hepatocellular carcinoma

Classification system	Proliferative class	Non-proliferative class
Lee (2004)	Cluster A	Cluster B
Boyault (2006)	G1, G2, G3	G4, G5, G6
Chiang (2008)	Proliferation	Interferon, Poly7, CTNNB1
Hoshida (2009)	S1, S2	S3
TCGARN (2017)	Immune high and intermediate	Immune excluded
Sia (2017)	iClust1, iClust3	iClust2
Clinical features	Poor clinical outcome, high AFP, HBV, frequent vascular invasion	Improved clinical outcome, low AFP, HCV, low vascular invasion
Histological features	Poorly differentiated	Well differentiated
Molecular alterations	Chromosomal instability, TP53 mutations, FGF19 amplification	Chromosomal stability, CTNNB1 mutations, TERT promoter mutations

List of abbreviations: AFP, alpha-fetoprotein; HBV, hepatitis B virus; HCV, hepatitis C virus

Figure legends

Figure 1. Microscopic features of a typical HCC. (A) Non-neoplastic hepatocytes (lower half) and HCC tumor cells (upper half) are separated by a fibrous capsule. (B) Trabecular pattern HCC with steatosis. (C) Pseudoglandular pattern HCC with cholestasis. (Hematoxylin-Eosin stain, original magnification x100 (A, B), x200 (C))

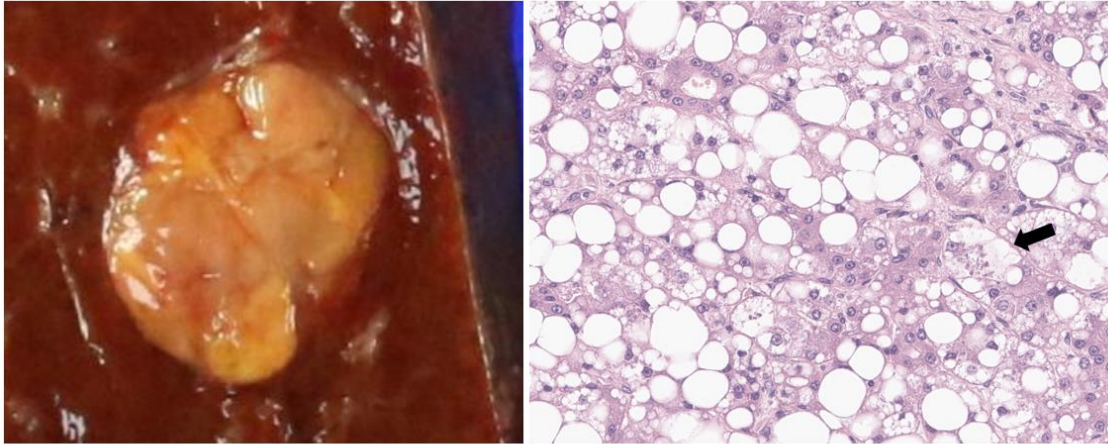


A

B

C

Figure 2. Steatohepatic HCC. (A) The tumor demonstrates a yellow hue on macroscopy reflecting the lipid component. (B) High power magnification showing the diffuse steatosis of tumor cells, tumor cell ballooning (arrow), some inflammatory cells, and pericellular fibrosis (Hematoxylin-Eosin stain, original magnification x400).



A

B

Epub

Figure 3. Clear cell HCC. Most of the tumor cells demonstrate clear cytoplasm due to glycogen accumulation. (Hematoxylin-Eosin stain, original magnification x200).

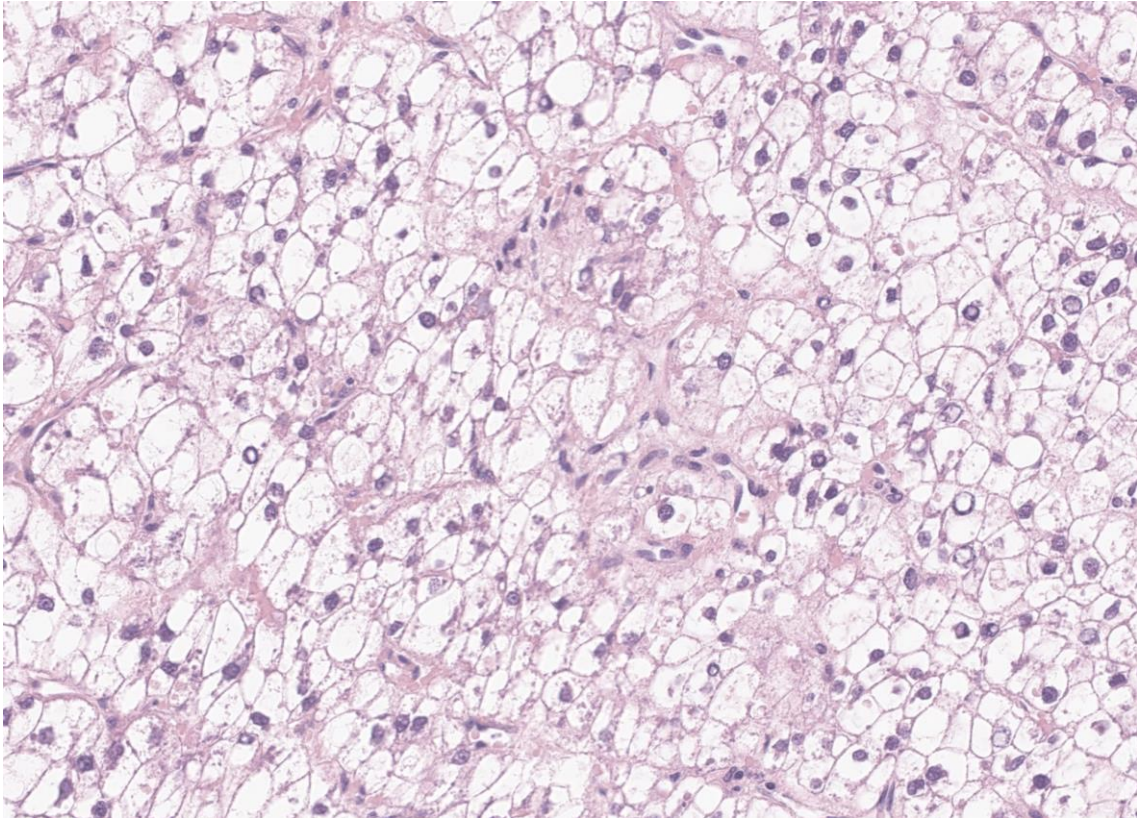


Figure 4. Macrotrabecular-massive HCC (A) and vessels-encapsulating-tumor clusters (VETC) pattern (B). (A) Macrotrabecular-massive HCC demonstrating thick tumor cell trabeculae, of more than 10-cell thickness (Hematoxylin-Eosin stain, original magnification x100). (B) CD34 immunostain highlighting the VETC pattern, where the CD34-positive endothelial cells completely surround tumor cell clusters.

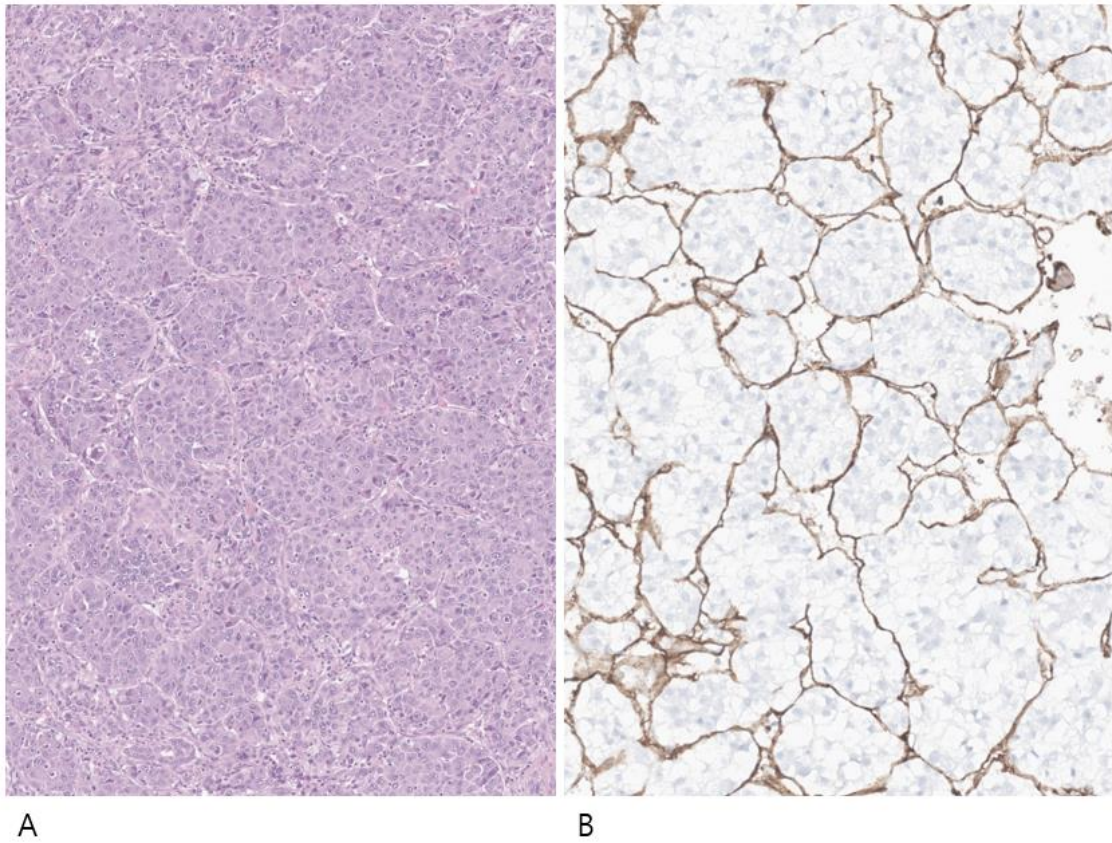


Figure 5. Scirrhou HCC. (A) The tumor appears as a firm, yellowish-white and lobulated mass on gross examination, mimicking an intrahepatic cholangiocarcinoma. (B) Dense intratumoral fibrosis is evident at low power magnification (Hematoxylin-Eosin stain, original magnification x40). (C) Immunohistochemical expression of cytokeratin 19 is seen in a few tumor cells.

