Review

Pathogenesis and management of metabolic dysfunction-associated steatohepatitis-related hepatocellular carcinoma: a narrative review

Running title: Pathogenesis and management of MASH-related HCC

Han Ah Lee*

Department of Internal Medicine, Chung-Ang University College of Medicine, Seoul, Korea

*Corresponding author: Han Ah Lee, Department of Internal Medicine, Chung-Ang University College of Medicine, 102, Heukseok-ro, Dongjak-gu, Seoul, 06973, Korea, E-mail: amelia86@naver.com

Abstract

Metabolic dysfunction-associated steatohepatitis (MASH) is increasingly recognized as a leading cause of hepatocellular carcinoma (HCC), the third-leading cause of cancer mortality worldwide, driven by the global obesity epidemic. Projected to become the primary cause of HCC by 2030, MASH-HCC presents unique clinical challenges. This review examines its clinical management, including surveillance strategies and treatment advances, and discusses prospects to overcome existing challenges. MASH-HCC accounts for 10–20% of HCC cases, particularly in Western countries, with a rising incidence due to obesity. Risk factors include cirrhosis, diabetes, obesity, alcohol, smoking, genetic polymorphisms (e.g., PNPLA3), and microbiome alterations. The pathogenesis involves fibrosis, immune dysfunction (e.g., T-cell impairment), and molecular changes. Prevention focuses on lifestyle modifications. Surveillance in patients with MASH

cirrhosis is crucial but is hindered by poor ultrasound sensitivity in obese patients, necessitating alternative methods. Treatment mirrors that of other HCC types, but comorbidities and potentially reduced efficacy of immunotherapy necessitate tailored approaches. MASH is becoming the leading cause of HCC, necessitating lifestyle interventions for prevention. Improved surveillance and early detection are critical but challenging due to obesity-related factors. Treatments align with those for other HCC types, but comorbidities and potential differences in immunotherapy efficacy due to T-cell dysfunction require careful consideration. Key needs include identifying molecular drivers in non-cirrhotic MASLD, developing preventive therapies, refining surveillance methods, and tailoring treatments. Trials should specifically report MASH-HCC outcomes to enable personalized therapies. Further research is needed to understand T-cell dysfunction, optimize immunotherapies, and identify predictive biomarkers.

Introduction

Background

Primary liver cancer ranks as the third leading cause of cancer-related deaths globally, with nearly 1 million new cases reported annually [1]. Hepatocellular carcinoma (HCC) accounts for approximately 90% of all primary liver cancers. It typically arises in the setting of chronic liver diseases, which may be due to hepatitis B virus (HBV), hepatitis C virus (HCV), alcohol-related liver disease, or metabolic dysfunction-associated steatotic liver disease (MASLD) [2]. MASLD is estimated to affect around 20–25% of the global population [3]. Metabolic dysfunction-associated steatohepatitis (MASH) is characterized by more than 5% steatosis, hepatocellular injury (such as "ballooning"), and inflammation, which may occur with or without fibrosis [4]. About 20% of individuals with MASLD develop MASH, which is strongly linked to rising rates of obesity, diabetes, and metabolic syndrome. As MASH progresses, it can lead to severe liver-related complications, including cirrhosis or liver failure, and significantly increases the risk of developing HCC [5].

In patients with MASH-related cirrhosis, the annual incidence of HCC is approximately 2% [6]. Moreover, MASH is the primary cause of HCC in patients who do not have cirrhosis [7]. MASH-related HCC accounts for 20% of HCC cases in the Western world and is projected to become the leading cause of HCC globally by 2030 [8]. The development of MASH-related HCC is characterized by unique mutational, immunological, and microenvironmental features. Although most cases of MASH-related HCC occur in patients with cirrhosis, 30–40% develop in those with advanced fibrosis but without cirrhosis. This suggests a distinct metabolic environment and the likely involvement of extrahepatic cancer drivers associated with metabolic syndrome [9, 10]. Unlike infections with HBV or HCV, MASH more frequently leads to HCC in the absence of cirrhosis, underscoring the need for strengthened surveillance and early detection [11].

Currently, MASH-HCC is managed similarly to other causes of HCC, employing strategies such as transplantation, resection, or locoregional therapies for early- or intermediate-stage disease [12]. MASH is the leading cause of HCC-related liver transplants in the USA; however, approximately 50% of patients undergo systemic therapy as their disease progresses, which includes both combination therapies and single-agent treatments with tyrosine kinase inhibitors or monoclonal antibodies [13]. Nevertheless, it remains uncertain whether immune-based therapies are as effective for non-viral HCC as they are for viral-related HCC [14].

Objectives

In this review, we examine the clinical management of MASH-HCC, focusing on surveillance strategies and recent advancements in treatment. We also discuss the customized application and outcomes of surgical, locoregional, and systemic therapies, examining future prospects and strategies to address current challenges.

Epidemiology

Approximately 10% (ranging from 1% to 38%) of all HCC cases are associated with MASLD, with higher rates (>20%) reported in studies from the USA, UK, India, Germany, and the Middle East. In contrast, lower estimates (1–2%) are reported from China and Japan [15]. The incidence of MASH-related HCC is expected to rise substantially as the obesity epidemic continues to expand [16]. Mathematical models predict a

significant increase in the incidence of MASH-HCC from 2016 to 2030, with projected rises of 47% in Japan, 82% in China, 88% in the UK, 117% in France, and 130% in the USA [17]. Compared to patients with HCC due to viral hepatitis (HBV or HCV) or alcohol-related liver disease, those with MASH-HCC typically have a lower male-to-female ratio (1.2:1), are generally 5–10 years older (mean age 73), and are more likely to have metabolic and cardiovascular comorbidities, such as type 2 diabetes mellitus and chronic vascular disease. Although the incidence of MASH-HCC is lower than that associated with active viral hepatitis, the increasing prevalence of MASLD, combined with improved treatments for viral hepatitis, is expected to increase both the proportion and rate of HCC attributed to MASLD [18, 19].

Risk factors

Liver cirrhosis

A study involving approximately 300,000 patients with MASLD reported an HCC incidence of 0.21 per 1,000 person-years, which is seven times higher than that observed in control individuals without liver disease— specifically, those free from viral hepatitis and with normal alanine aminotransferase levels [20]. The primary risk factor for MASH-HCC is cirrhosis, with incidence rates in cohorts of MASH cirrhosis estimated at about 2% per year, although these rates vary from 0.3% to 4.7% per year [6]. This variability can be attributed to differences in age, metabolic profiles, and the severity of liver decompensation. While HCC can also develop in MASH patients without cirrhosis, the overall incidence in this subgroup is low, ranging between 0.01% and 0.13% per year. It is even lower in the general MASLD population, underscoring the importance of assessing cirrhosis status as the primary risk stratifier for MASLD [21].

Diabetes

In cohort studies from both Europe and the US, type 2 diabetes has been identified as the strongest independent metabolic risk factor for the development of HCC. A retrospective study demonstrated that in patients with MASH-cirrhosis, the presence of diabetes mellitus (DM) was associated with a fourfold increase in the risk of developing HCC (hazard ratio [HR], 4.2; 95% confidence interval [CI], 1.2–14.2; P=0.02) [19].

Another large study in Europe, which included 136,703 patients with MASLD, found that among the 6,425 (4.7%) patients with advanced fibrosis, DM was the most significant risk factor for HCC [22]. Similarly, a study involving a US cohort of 271,906 MASLD patients, of whom 253 had HCC, reported a strong association between DM and HCC (adjusted HR, 2.77; 95% CI, 2.03–3.77) [23].

Obesity

In a large cohort study involving 296,707 patients, those diagnosed with MASLD and obesity did not show a statistically significant increase in HCC risk (P=0.06). However, the risk increased significantly, by 2.6 times, when obesity was accompanied by diabetes, hypertension, and hyperlipidemia [20]. Another recent study, which examined data from 98,090 MASLD patients with severe obesity, found that those who underwent bariatric surgery experienced a reduced risk of HCC. The adjusted HR was 0.48 (95% CI, 0.24–0.89) [24]. Although numerous studies have explored the link between obesity and elevated HCC risk, most have not sufficiently evaluated the presence of MASLD or MASH.

Alcohol

The impact of mild to moderate alcohol consumption on the development of HCC in patients with MASLD is still unclear, as research has produced inconsistent findings. A cohort study in Korea examined the relationship between mild to moderate alcohol intake and the progression of non-invasive fibrosis scores in 58,927 adults with MASLD who initially had low fibrosis scores over a median period of 4.9 years [25]. Of these participants, 5,303 (9%) progressed from low to intermediate or high fibrosis scores. Moderate drinkers were more likely to experience increased fibrosis compared to nondrinkers, with an HR of 1.29 (95% CI, 1.23). Another study indicated that even mild drinking habits increased the risk of carcinogenesis in patients with MASH-associated cirrhosis, presenting an HR of 3.8 (95% CI, 1.6–8.9; P=0.002); however, this study focused solely on patients with decompensated liver disease [26]. Additionally, a recent multivariate analysis of patients with biopsy-proven MASLD across various stages of fibrosis revealed that consuming less than 20 g of alcohol per day heightened the risk of HCC, especially in those with advanced F3–4 fibrosis, with a relative risk (RR) of 4.83 (P=0.04) [27].

Smoking

Smoking is generally associated with an increased risk of HCC; however, its specific impact on MASLD has not been thoroughly investigated [28].

Coffee

Coffee is rich in antioxidants, including phenolic compounds such as chlorogenic, caffeic, ferulic, and coumaric acids, along with melanoidins and diterpenes such as cafestol and kahweol. These compounds have shown inhibitory effects on the development of HCC [29, 30]. Additionally, the beneficial effects of coffee in preventing HCC may be partially attributed to its role in lowering the risk of type 2 DM, which is a known risk factor for HCC [31].

Antidiabetics

Metformin inhibits the mammalian target of the rapamycin pathway, which plays a role in cell proliferation by activating AMP-activated protein kinase (AMPK) [32]. It also inhibits angiogenesis, disrupts the cell cycle, and induces apoptosis independently of p53 [33]. Additionally, metformin promotes moderate weight loss, mitigates the effects of hyperinsulinemia on the cell cycle and inflammation, and improves liver biochemistry and histology in patients with MASLD [34, 35]. Research has explored the impact of antidiabetic medications on HCC risk, recognizing diabetes as a significant risk factor. A recent study demonstrated that effective glycemic control was associated with a 31% reduced risk of HCC in patients with MASLD and DM [36]. The study also found that metformin use led to a 20% decrease in HCC risk, whereas insulin use, particularly when combined with other oral antidiabetic medications, increased the risk by 1.6 to 1.7 times. However, a database study of 18,080 MASLD patients without cirrhosis, monitored over an average of 6.3 years, showed no link between metformin use and HCC risk [37]. In a recent nationwide cohort, patients with MASLD and DM who used sodium-glucose cotransporter-2 inhibitors had significantly lower risks of liver and non-liver complications compared to users of other antidiabetic medications, with HRs ranging from 0.76 to 0.97. The risk was further reduced when metformin was also used, with HRs between 0.58 and 0.79 [38].

Statins

Statins exhibit a range of anticancer effects that go beyond their ability to lower cholesterol. They inhibit key oncogenic drivers including MYC, AKT, Rho-dependent kinase, and extracellular signal-regulated kinase 1 and 2 [35, 39, 40]. Additionally, statins activate protective liver pathways such as AMPK and p38-MAPK, and promote apoptosis through a p53-dependent mechanism [41, 42]. These drugs have also been linked to anticarcinogenic effects. A database study from Taiwan involving 18,080 MASLD patients demonstrated an inverse relationship between statin use and HCC, with an odds ratio (OR) of 0.29 (95% CI, 0.12–0.68) [37]. In a retrospective case-control study of 102 MASLD patients, including 34 HCC cases, statins were found to be protective against HCC (OR, 0.20; 95% CI, 0.07–0.60) [43]. Another recent retrospective study showed that statin use significantly and dose-dependently reduced the risk of HCC in patients with NASH cirrhosis [44]. However, a study involving 458 MASLD patients with advanced fibrosis did not find such an association [45]. The uncontrolled and retrospective nature of these studies limits the ability to definitively interpret their findings on the chemopreventive benefits of statins, making it inappropriate to recommend them solely for the prevention of HCC.

Pathogenesis

Liver fibrosis

Approximately 80% of MASLD patients do not develop NASH, prompting research efforts to focus on identifying the factors that differentiate those with inflammation, cell injury, and fibrosis (MASH) from those exhibiting simple steatosis. A critical factor in understanding the progression to MASH is lipotoxicity, which involves hepatocellular injury resulting from disrupted fat metabolism [46]. Lipotoxicity is triggered by various factors, including increased fatty acid delivery to the liver, insulin resistance, and inflammatory signals from dysfunctional adipose tissue [47]. This condition leads to cellular stress, oxidative damage, inflammasome activation, and ultimately, cell death in hepatocytes [48]. These damaging responses are linked to pre-malignant changes, such as oxidative DNA damage and mutations in metabolism-related genes such as *FOXO1*, *CIDEB*, and *GPAM*. Although these genes may help protect hepatocytes from lipotoxicity, they also

elevate the risk of malignancy [49, 50].

To repair hepatocellular injuries in MASH, developmental pathways such as YAP–TAZ, Notch, and Hedgehog signaling are reactivated in hepatocytes. This reactivation leads to cell proliferation, inflammation, and potentially cancer [51, 52]. In advanced MASH, there is a marked decline in hepatocyte proliferation and regenerative capacity. These dysregulated cells exacerbate inflammation and fibrosis [53]. Consequently, this hepatocellular damage fosters a pro-inflammatory environment, perpetuating chronic inflammation and impacting various immune cell types.

The stage of hepatic fibrosis in MASH is a critical determinant of clinical outcomes, as it can progress to cirrhosis and liver failure, and create conditions conducive to cancer development [54]. This process involves the activation or transdifferentiation of resident hepatic stellate cells (HSCs) into fibrogenic, proliferating myofibroblasts, which leads to the accumulation of extracellular matrix or scar tissue. Advanced single-cell sequencing has revealed significant heterogeneity among HSCs in MASH, although the functional implications of this diversity are not yet clear [55]. The exact mechanisms by which MASH-HCC develops without cirrhosis remain poorly understood, but they are likely related to fibrosis. The accumulation of extracellular matrix increases liver stiffness, which can facilitate the emergence and growth of tumor cells [56]. This scar matrix also acts as a reservoir for growth factors that may support the survival of pre-neoplastic hepatocytes, thereby promoting tumor initiation or progression. Additionally, HSCs possess immunoregulatory properties that contribute to the liver's immune tolerance, potentially affecting its response to checkpoint blockade therapies [57].

Angiogenesis is implicated in both MASH and potentially MASH-HCC. Increased CD34 expression in new blood vessels has been observed in previous studies involving both humans and rodents, indicating enhanced vascularization [58]. Vascular endothelial growth factor (VEGF), a crucial angiogenic signal, shows elevated levels in experimental MASH models. Inhibiting VEGF leads to reduced vascularization, inflammation, and steatosis [59].

The impact of treatments targeting MASH on the risk of MASH-HCC has yet to be determined;

however, a decrease in HCC risk has been noted in MASH patients following bariatric surgery, indicating that future medical interventions for MASH could potentially lower the incidence of HCC [60]. Nonetheless, it remains uncertain whether advanced liver fibrosis continues to carry an inherent risk of cancer even if the fibrosis subsequently regresses.

Immune system

The immune system plays a major role in both MASLD and HCC, and distinct immunogenomic classifications have been identified [61]. MASH is characterized by inflammatory responses in the liver, which are pivotal in its progression to fibrosis, cirrhosis, or HCC [62]. Both innate and adaptive immune mechanisms significantly contribute to hepatic inflammation in MASH. Resident Kupffer cells and the recruitment of leukocytes, including neutrophils, monocytes, NK cells, and NKT cells, promote inflammation through the release of cytokines, chemokines, and reactive oxygen species. Elevated levels of CD4+ T helper cells, particularly the TH1 and TH17 subsets, have been observed in the livers of mice with MASH [63]. Although T cells exhibit anti-tumorigenic properties, the depletion of CD8+ T cells accelerates tumor growth in MASH-driven HCC models. Similarly, the depletion of CD4+ T cells promotes tumor growth, impacting the efficacy of immune-based therapies [64].

The disruption of the immune system in MASH and MASH-HCC has been linked to the response to immunotherapies. Both adaptive and innate immune cells, including CD4+ T cells, metabolically activated CD8+ T cells, platelets, and dendritic cells, play a role in shaping the liver microenvironment as MASH progresses to HCC [65, 66]. Neutrophils, in particular, are involved in the transition from fatty liver to steatohepatitis. They contribute to an immunosuppressive environment through the production of extracellular traps and PDL1 signaling, which leads to CD8+ T cell exhaustion and affects the response to immunotherapy [67, 68]. In MASLD, impaired antigen-specific T-cell function has been observed, partially due to macrophage activity [69]. In advanced HCC, the infiltration of CCR2+ and CX3CR1+ macrophages is linked to non-responsiveness to immune-checkpoint inhibition. Conversely, pro-inflammatory PDL1-expressing CXCL10+ macrophages can drive treatment response. Recent studies indicate that T cells lose functionality in MASLD, which contributes to poor responses to immune checkpoint inhibitor (ICI) therapy

[70]. Approaches such as neutrophil reprogramming with CXCR2 antagonists have shown promise in enhancing the effectiveness of ICI therapy in MASH-HCC models by increasing dendritic cell activity and CD8+ T cell numbers [68].

In two notable studies involving both mice and humans, the presence of CD8+PD1+ T cells in the liver increased as MASH progressed. These cells are in an auto-aggressive state, characterized by liver-resident CD8+PD1+CD103+ T cells that, despite being exhausted, display an activated phenotype and express high levels of cytokines such as TNF, CCL2, IL-10, and granzyme B [71, 72]. In MASH-HCC mouse models treated with immunotherapy, these CD8+PD1+ cells exhibited minimal changes in their transcriptomes and proteomes, yet they increased in size over time. This growth contributed to heightened liver inflammation, hepatocyte death, and oncogenic signaling [72]. Instead of eliminating HCC, these cells became dysfunctional in tumor surveillance and even promoted tumor growth. This dysfunction resulted in a lack of response to ICIs in therapeutic settings and accelerated HCC development in preventive scenarios. Similar characteristics of CD8+ T cells have been observed in human MASH-HCC, indicating that peritumoral and intratumoral CD8+PD1+ T cells could potentially serve as predictors of treatment success or resistance to ICIs. Understanding the immune microenvironment is essential for identifying the most effective therapies in future research.

Microbiome

The gut microbiome plays a crucial role in influencing altered liver responses in MASH by affecting hepatic bile acid metabolism and facilitating the translocation of gut-derived signals through an increasingly permeable gut lining [73]. Throughout all stages of NASH, the gut–liver axis remains active, with interactions between liver damage, regeneration, and heightened gut permeability exacerbating inflammatory, pro-fibrogenic, and pro-carcinogenic pathways [48]. This permeability defect allows for both direct (e.g., bacterial presence) and indirect (e.g., bacterial metabolites) interactions between the gut microbiome and the liver, which in turn impact liver metabolism and contribute to the progression of MASH and HCC.

The gut microbiome has been identified as a key factor in triggering MASLD, driving liver steatosis

by enhancing energy harvest, monosaccharide absorption, and abnormal acetate production [74]. A dysbiotic, leaky gut permits the translocation of pathogen-associated and danger-associated molecular patterns into the liver, activating immune cells and Toll-like receptors, which in turn trigger pro-inflammatory and fibrotic pathways [75]. In mice, disruption of the gut vascular barrier by the microbiota is seen as a precursor to NASH [76]. Additionally, inflammatory cells from the gut may migrate to the liver, contributing to bacterial translocation. Several bacterial species, such as Proteobacteria, Enterobacteriaceae, and Escherichia, are associated with MASLD in humans, and levels of Bacteroides are elevated in MASH patients [76, 77]. Treatment with non-absorbable antibiotics, such as rifaximin, has shown potential in improving liver function, underscoring the significant role of the gut microbiome in MASH pathogenesis [78].

4) Molecular alterations

Several single-nucleotide polymorphisms (SNPs) associated with abnormal lipid metabolism in hepatocytes have been linked to an increased risk of MASH and progression to HCC. One of the most well-known SNPs is rs738409 in the *PNPLA3* gene, which encodes the patatin-like phospholipase domain-containing protein 3. This variant interferes with the breakdown of lipid droplets in hepatocytes, leading to decreased triglyceride lipolysis and promoting hepatic steatosis. As a result, it is associated with more than a 2-fold increased risk of MASH and a 2.2-fold higher risk of progressing to MASH HCC compared to those without the variant [79]. Another significant SNP, rs58542926 in the *TM6SF2* gene, plays a role in regulating liver fat metabolism and increases hepatic triglyceride content. This variant is linked to a 1.6-fold increased risk of MASH and a 1.9-fold higher risk of MASH HCC [80].

Additionally, an SNP near the *MBOAT7* gene is associated with increased hepatic triglyceride levels and occurs twice as frequently in patients with MASH-HCC compared to those with MASLD alone [81]. A loss-of-function variant in the GCKR gene, which encodes the glucokinase regulator, leads to increased de novo lipogenesis and insulin resistance. This variant is linked to a 1.5-fold increased risk of MASH and a 1.8-fold higher risk of MASH-HCC [82]. A polygenic risk score that incorporates these four SNPs has been suggested for HCC risk stratification in patients of European ancestry with NASH cirrhosis. This score has proven to be a more accurate predictor of HCC development than individual SNPs ($P<10^{-13}$) [83].

MASH-HCC is often associated with an increased presence of ACVR2A and TP53 mutations, as well as the proliferative class S1-WNT/TGFβ [84]. A distinct mutational signature, termed MutSigNASH-HCC, has been identified in 25% of MASH-HCC patients, compared to only 2% in those with other causes. This signature is characterized by a higher frequency of C>T and C>A transitions [85]. Furthermore, patients with MASH-HCC exhibit higher levels of hepatic oxidative DNA damage than those with other etiologies, a phenomenon that correlates with a diminished DNA damage response in experimental models [49]. Additionally, epigenetic events that suppress the transcription of genes involved in bile and fatty acid metabolism, while activating proliferative pathways, have been implicated in MASH-HCC. Experimental models have shown that epigenetic reprogramming can reverse hepatocarcinogenesis [86].

A diagram of the pathogenesis of hepatocellular carcinoma associated with metabolic dysfunction-related steatohepatitis is shown in Fig. 1.

Clinical management

Prevention

Several observational, retrospective, population-based studies have suggested that metformin, statins, coffee, and aspirin might contribute to the prevention of HCC, regardless of the underlying liver disease etiology [87, 88]. Due to its generally favorable benefit-to-risk ratio, current guidelines endorse the consumption of coffee for individuals with chronic liver disease [89, 90]. However, other agents have not demonstrated sufficient efficacy to be recommended for HCC prevention, and most studies related to this have not been conducted in well-defined populations with MASLD.

For the prevention of MASH-HCC, the American Association for the Study of Liver Diseases (AASLD), the European Association for the Study of the Liver (EASL), and the Korean Association for the Study of Liver Diseases (KASL) recommend combining a hypocaloric or Mediterranean diet with moderate-intensity exercise to achieve and maintain weight loss, as outlined in their practice guidelines [89–91]. Additionally, a large multinational cohort study has demonstrated that physical activity is associated with a

reduced risk of HCC [92]. Although there is no direct evidence currently available that weight loss decreases the risk of MASH-HCC, observational studies indicate that weight loss may reverse steatosis and potentially fibrosis in patients with MASH, thereby suggesting a possible benefit of weight loss in reducing the risk of HCC [93, 94].

HCC surveillance

The clinical practice guidelines from the AASLD, EASL, and KASL recommend semiannual surveillance for HCC using abdominal ultrasound, with or without α-fetoprotein testing, for all patients with cirrhosis, regardless of the underlying cause. However, only two studies have specifically assessed the potential benefits of such surveillance in patients with MASLD-related cirrhosis.

However, a previous study found no significant association between surveillance and the applicability of curative treatment (45.5% versus 51.5%; P=0.72) [95].

Data specifically focusing on patients with MASLD are important, as this group exhibits unique characteristics that pose challenges to traditional HCC surveillance methods. Notably, about one-third of MASLD-HCC cases arise in individuals without cirrhosis, suggesting that these patients are often excluded from the at-risk populations typically targeted for surveillance [21]. Furthermore, at the time of their HCC diagnosis, patients with MASLD are generally less likely to have been previously diagnosed with liver disease or cirrhosis, which likely contributes to their lower rates of surveillance utilization [96]. A meta-analysis revealed that a significantly smaller proportion of patients with MASLD-HCC (32.8%, 95% CI, 12.0–63.7) underwent surveillance compared to patients with HCC from other causes (55.7%, 95% CI, 24.0–83.3; P<0.0001) [97].

Second, patients with MASH are more likely to experience inadequate ultrasound visualization and surveillance failure, leading to a higher rate of late-stage HCC diagnoses even when surveillance is performed [98, 99]. This suggests that the sensitivity of ultrasound-based surveillance in patients with MASH may be lower than the 63% observed in those with HCC from other causes [100]. This finding underscores the need for alternative imaging methods, such as computed tomography or magnetic resonance imaging, and blood-

based biomarker strategies for this group [101].

Treatment for MASH-HCC

Patients with MASH-HCC often present with comorbidities, such as cardiovascular disease, which can restrict their access to curative treatments, especially surgery [102, 103]. However, a systematic review has shown that despite having more comorbidities and larger tumors at diagnosis, the allocation of treatments for MASLD patients is similar to that for other patients [97]. Moreover, when severe comorbidities are excluded, the outcomes following curative and locoregional treatments for MASH-HCC are comparable to, or even better than, those observed in non-MASH patients. Lastly, immunotherapies may be less effective in non-viral HCC cases, such as MASH-HCC, due to impairments in the immune system [72].

Surgery: Patients with MASLD face a higher risk of intra-operative complications and poorer post-surgical outcomes, largely due to the increased prevalence of metabolic syndrome comorbidities. Obesity and type 2 diabetes have been linked to lower survival rates in cancer patients, including those receiving surgical treatments [104, 105]. Research indicates that patients with MASH-HCC are more likely to suffer from hypertension, hyperlipidemia, and ischemic heart disease compared to those with other causes of HCC, all factors that heighten the risk of post-surgical morbidity and complications [102]. Furthermore, the degree of liver steatosis may correlate with poorer surgical outcomes [106].

However, a systematic review and meta-analysis of 14 studies, which included 7,226 HCC patients—approximately 20% of whom had MASH-HCC—demonstrated that patients with MASH-HCC experienced improved disease-free survival (HR, 0.81) and overall survival (HR, 0.78) compared to those with other causes [107]. Another meta-analysis corroborated these results, suggesting that the better outcomes in MASH-HCC patients might be due to the absence of cirrhosis in many cases and the exclusion of those with severe comorbidities from surgical interventions [108].

Liver transplantation: An analysis of the United Network for Organ Sharing (UNOS) registry from 2002 to 2012 revealed that patients with MASH-HCC had significantly better post-transplant survival outcomes (HR, 0.69; 95% CI, 0.63–0.77) and a lower risk of graft failure (HR, 0.76; 95% CI, 0.69–0.83)

compared to those with other causes of HCC. This was despite a higher prevalence of diabetes and cardiovascular disease in the MASH-HCC group [109]. In contrast, data from the European Liver Transplant Registry showed no statistically significant differences in post-transplant survival or graft survival between patients with HCC, regardless of MASLD status. However, there were differences in the causes of mortality [110]. While some single-center studies suggest that patients with MASLD may have a higher risk of post-transplant complications, the overall evidence indicates similar post-transplant survival rates between patients with MASLD and those with other etiologies of HCC [111].

Locoregional therapies: Current evidence on the efficacy of locoregional therapies for MASH-HCC is limited. However, a study using the SEER-Medicare database showed similar overall survival rates following radiofrequency ablation in patients with MASH-HCC compared to those with other HCC etiologies [112]. Additionally, a propensity score-matched study that included patients undergoing transarterial chemoembolization (TACE) revealed no significant differences in time-to-progression (13.0 vs. 8.5 months; P=0.25) or overall survival (23.2 vs. 28.0 months; P=0.48) between patients with and without MASLD [113]. Another study comparing MASLD-HCC and HBV-related HCC patients treated with transarterial radioembolization (TARE) also found no significant differences in treatment-related adverse events or overall survival [114]. These results indicate that TACE and TARE are likely safe and effective treatments for patients with MASH-HCC, yielding comparable outcomes across different etiologies.

Systemic therapies: Phase III studies of systemic therapies in advanced HCC have predominantly involved patients with compensated liver disease. However, the etiology of liver disease has not been a consideration in treatment decisions or trial designs. Typically, studies report efficacy based on stratification factors such as etiology, often categorized as HBV, HCV, or "non-viral." The "non-viral" category includes alcohol-related disease, MASH, and other causes (Table 1).

Currently, several agents are approved for the first- and second-line treatment of advanced HCC. These can be broadly categorized into two groups: multi-kinase VEGFR-targeting small molecules and VEGFR2 monoclonal antibody approaches, as well as immunotherapy-based approaches. Regarding overall survival, the efficacy of the first group does not significantly vary based on the etiology of HCC, as

evidenced by similar HRs for overall survival in the study versus control arms. This trend is also generally observed in secondary endpoints, such as progression-free survival and objective response rates (ORRs).

Unlike previous treatments, ICIs have not only demonstrated a survival benefit but have also achieved significant response rates with durable responses lasting over 20 months. There is growing interest in evaluating clinical characteristics as markers of benefit, especially those associated with distinct pathogenic pathways and immune profiles linked to different HCC etiologies. Two studies have raised questions about the effectiveness of immunotherapies in metabolic-associated steatohepatitis-HCC (MASH-HCC) compared to viral-related HCC [72, 115]. However, none of the phase III randomized controlled trials (RCTs) in advanced HCC have reported the percentage of patients with MASH-HCC. Consequently, indirect analysis of survival effects by etiology has been limited to non-viral HCC cases. A meta-analysis of three RCTs (IMbrave150, CheckMate 459, and Keynote-240) indicated that patients with viral-related HCC responded better to immunotherapies (HR, 0.64; 95% CI, 0.50–0.83) than those with non-viral-related HCC (HR, 0.92; 95% CI, 0.77–1.11; P=0.2) [115]. Following the publication of a subgroup analysis from the COSMIC-312 trial, a meta-analysis of four RCTs confirmed a significant difference in efficacy (P=0.01) [116]. When the HIMALAYA trial, which assessed a combination of two ICIs, was included in the meta-analysis (five RCTs), the difference remained significant, albeit less pronounced (P=0.046) [15]. These findings suggest that immunotherapies may be more effective in viral-related HCC than in other etiologies, supporting observations that MASH-HCC tumors have dysfunctional T cells, which may limit the effectiveness of ICIs [72].

However, these subgroup analyses are not statistically definitive and do not account for other prognostic factors. The term "non-viral etiologies" includes MASH-related, alcohol-related, idiopathic, and other metabolic causes, which complicates the analysis. These findings suggest that future studies should stratify participants based on etiology; however, dedicated prospective studies are necessary to determine the specific role of etiology. Although MASH-HCC is biologically distinct, the current clinical approaches remain consistent with those used for other non-viral etiologies, including alcohol-related HCC. Future trials should specifically identify cases of MASH-related HCC to better understand the impact of immunotherapies on the

survival of this subgroup.

Conclusion

MASH is a significant global health issue and is projected to become the leading cause of HCC by 2030. The progression from MASH to HCC is influenced by molecular changes, the stage of fibrosis, the immune microenvironment, and the microbiome. Lifestyle changes are crucial for preventing MASLD progression, and surveillance in patients with MASH cirrhosis enables earlier detection and improves survival. Currently, MASH-HCC is managed similarly to other HCC etiologies, but comorbidities such as obesity and diabetes can complicate treatment.

Key unmet needs include identifying the molecular drivers of HCC in non-cirrhotic MASLD and developing preventive therapies. There is also a need for improved surveillance methods, particularly alternatives to ultrasound for obese patients, and for refining the selection of surgical candidates. It is crucial to report MASH-HCC outcomes separately in trials to facilitate better analysis; thus, it is recommended that MASH-HCC be specifically identified in clinical trials to enable more effective, personalized treatments. Additionally, further studies are required to understand MASH-HCC-related T-cell dysfunction and to identify biomarkers that predict treatment responses.

ORCID

Han Ah Lee: https://orcid.org/0000-0003-4082-1121

Conflict of interest

No potential conflict of interest relevant to this article was reported.

Data availability

Not applicable.

Supplementary materials

Not applicable.

References

1. Kim DY. Changing etiology and epidemiology of hepatocellular carcinoma: Asia and worldwide. J Liver Cancer 2024;24:62-70.

2. 2022 KLCA-NCC Korea practice guidelines for the management of hepatocellular carcinoma. J Liver Cancer 2023;23:1-120.

3. Lee HH, Lee HA, Kim EJ, Kim HY, Kim HC, Ahn SH, Lee H, et al. Metabolic dysfunction-associated steatotic liver disease and risk of cardiovascular disease. Gut 2024;73:533-540.

4. Yeh ML, Yu ML. From nonalcoholic steatohepatitis, metabolic dysfunction-associated fatty liver disease, to steatotic liver disease: Updates of nomenclature and impact on clinical trials. Clin Mol Hepatol 2023;29:969-972.

5.	Anstee QM, Reeves HL, Kotsiliti E, Govaere O, Heikenwalder M. From NASH to HCC: current concepts and future challenges. Nat Rev Gastroenterol Hepatol 2019;16:411-428.

6.	Huang DQ, El-Serag HB, Loomba R. Global epidemiology of NAFLD-related HCC: trends, predictions, risk factors and prevention. Nat Rev Gastroenterol Hepatol 2021;18:223-238.

7.	Kim GA, Moon JH, Kim W. Critical appraisal of metabolic dysfunction-associated steatotic liver disease: Implication of Janus-faced modernity. Clin Mol Hepatol 2023;29:831-843.

8.	Le MH, Le DM, Baez TC, Dang H, Nguyen VH, Lee K, Stave CD, et al. Global incidence of adverse clinical events in non-alcoholic fatty liver disease: A systematic review and meta-analysis. Clin Mol Hepatol 2024;30:235-246.

9.	Han JW, Sohn W, Choi GH, Jang JW, Seo GH, Kim BH, Choi JY. Evolving trends in treatment patterns for hepatocellular carcinoma in Korea from 2008 to 2022: a nationwide population-based study. J Liver Cancer 2024.

10.	Piscaglia F, Svegliati-Baroni G, Barchetti A, Pecorelli A, Marinelli S, Tiribelli C, Bellentani S. Clinical patterns of hepatocellular carcinoma in nonalcoholic fatty liver disease: A multicenter prospective study. Hepatology 2016;63:827-838.

11.	Stine JG, Wentworth BJ, Zimmet A, Rinella ME, Loomba R, Caldwell SH, Argo CK. Systematic review with meta-analysis: risk of hepatocellular carcinoma in non-alcoholic steatohepatitis without cirrhosis compared to other liver diseases. Aliment Pharmacol Ther 2018;48:696-703.

12.	Kim DH. Combination of interventional oncology local therapies and immunotherapy for the treatment of hepatocellular carcinoma. J Liver Cancer 2022;22:93-102.

13.	Llovet JM, Castet F, Heikenwalder M, Maini MK, Mazzaferro V, Pinato DJ, Pikarsky E, et al. Immunotherapies for hepatocellular carcinoma. Nat Rev Clin Oncol 2022;19:151-172.

14.	Llovet JM, Heikenwalder M. Atezolizumab Plus Bevacizumab in Advanced HCC: Efficacy in NASH-Specific Etiology. Gastroenterology 2023;165:1308-1310.

15.	Llovet JM, Willoughby CE, Singal AG, Greten TF, Heikenwälder M, El-Serag HB, Finn RS, et al. Nonalcoholic steatohepatitis-related hepatocellular carcinoma: pathogenesis and treatment. Nat Rev Gastroenterol Hepatol 2023;20:487-503.

16.   Dyson J, Jaques B, Chattopadyhay D, Lochan R, Graham J, Das D, Aslam T, et al. Hepatocellular cancer: the impact of obesity, type 2 diabetes and a multidisciplinary team. J Hepatol 2014;60:110-117.

17.   Estes C, Anstee QM, Arias-Loste MT, Bantel H, Bellentani S, Caballeria J, Colombo M, et al. Modeling NAFLD disease burden in China, France, Germany, Italy, Japan, Spain, United Kingdom, and United States for the period 2016-2030. J Hepatol 2018;69:896-904.

18.   White DL, Kanwal F, El-Serag HB. Association between nonalcoholic fatty liver disease and risk for hepatocellular cancer, based on systematic review. Clin Gastroenterol Hepatol 2012;10:1342-1359.e1342.

19.   Yang JD, Ahmed F, Mara KC, Addissie BD, Allen AM, Gores GJ, Roberts LR. Diabetes Is Associated With Increased Risk of Hepatocellular Carcinoma in Patients With Cirrhosis From Nonalcoholic Fatty Liver Disease. Hepatology 2020;71:907-916.

20.   Kanwal F, Kramer JR, Mapakshi S, Natarajan Y, Chayanupatkul M, Richardson PA, Li L, et al. Risk of Hepatocellular Cancer in Patients With Non-Alcoholic Fatty Liver Disease. Gastroenterology 2018;155:1828-1837.e1822.

21.   Mittal S, El-Serag HB, Sada YH, Kanwal F, Duan Z, Temple S, May SB, et al. Hepatocellular Carcinoma in the Absence of Cirrhosis in United States Veterans is Associated With Nonalcoholic Fatty Liver Disease. Clin Gastroenterol Hepatol 2016;14:124-131.e121.

22.   Alexander M, Loomis AK, van der Lei J, Duarte-Salles T, Prieto-Alhambra D, Ansell D, Pasqua A, et al. Risks and clinical predictors of cirrhosis and hepatocellular carcinoma diagnoses in adults with diagnosed NAFLD: real-world study of 18 million patients in four European cohorts. BMC Med 2019;17:95.

23.   Kanwal F, Kramer JR, Li L, Dai J, Natarajan Y, Yu X, Asch SM, et al. Effect of Metabolic Traits on the Risk of Cirrhosis and Hepatocellular Cancer in Nonalcoholic Fatty Liver Disease. Hepatology 2020;71:808-819.

24.   Rustgi VK, Li Y, Gupta K, Minacapelli CD, Bhurwal A, Catalano C, Elsaid MI. Bariatric Surgery Reduces Cancer Risk in Adults With Nonalcoholic Fatty Liver Disease and Severe Obesity. Gastroenterology 2021;161:171-184.e110.

25.   Chang Y, Cho YK, Kim Y, Sung E, Ahn J, Jung HS, Yun KE, et al. Nonheavy Drinking and Worsening of Non-invasive Fibrosis Markers in Nonalcoholic Fatty Liver Disease: A Cohort Study. Hepatology 2019;69:64-75.

26.   Ascha MS, Hanouneh IA, Lopez R, Tamimi TA, Feldstein AF, Zein NN. The incidence and risk factors of hepatocellular carcinoma in patients with nonalcoholic steatohepatitis. Hepatology 2010;51:1972-1978.

27.   Kimura T, Tanaka N, Fujimori N, Sugiura A, Yamazaki T, Joshita S, Komatsu M, et al. Mild drinking habit is a risk factor for hepatocarcinogenesis in non-alcoholic fatty liver disease with advanced fibrosis. World J Gastroenterol 2018;24:1440-1450.

28.   Abdel-Rahman O, Helbling D, Schöb O, Eltobgy M, Mohamed H, Schmidt J, Giryes A, et al. Cigarette smoking as a risk factor for the development of and mortality from hepatocellular carcinoma: An updated systematic review of 81 epidemiological studies. J Evid Based Med 2017;10:245-254.

29.   Cavin C, Holzhaeuser D, Scharf G, Constable A, Huber WW, Schilter B. Cafestol and kahweol, two coffee specific diterpenes with anticarcinogenic activity. Food Chem Toxicol 2002;40:1155-1163.

30.   Majer BJ, Hofer E, Cavin C, Lhoste E, Uhl M, Glatt HR, Meinl W, et al. Coffee diterpenes prevent the genotoxic effects of 2-amino-1-methyl-6-phenylimidazo[4,5-b]pyridine (PhIP) and N-nitrosodimethylamine in a human derived liver cell line (HepG2). Food Chem Toxicol 2005;43:433-441.

31.   Huxley R, Lee CM, Barzi F, Timmermeister L, Czernichow S, Perkovic V, Grobbee DE, et al. Coffee, decaffeinated coffee, and tea consumption in relation to incident type 2 diabetes mellitus: a systematic review with meta-analysis. Arch Intern Med 2009;169:2053-2063.

32.   Zheng L, Yang W, Wu F, Wang C, Yu L, Tang L, Qiu B, et al. Prognostic significance of AMPK activation and therapeutic effects of metformin in hepatocellular carcinoma. Clin Cancer Res 2013;19:5372-5380.

33.   Buzzai M, Jones RG, Amaravadi RK, Lum JJ, DeBerardinis RJ, Zhao F, Viollet B, et al. Systemic treatment with the antidiabetic drug metformin selectively impairs p53-deficient tumor cell growth. Cancer Res 2007;67:6745-6752.

34.   Forslund K, Hildebrand F, Nielsen T, Falony G, Le Chatelier E, Sunagawa S, Prifti E, et al. Disentangling type 2 diabetes and metformin treatment signatures in the human gut microbiota. Nature 2015;528:262-266.

35.   Cao Z, Fan-Minogue H, Bellovin DI, Yevtodiyenko A, Arzeno J, Yang Q, Gambhir SS, et al. MYC phosphorylation, activation, and tumorigenic potential in hepatocellular carcinoma are regulated by HMG-CoA reductase. Cancer Res 2011;71:2286-2297.

36.   Kramer JR, Natarajan Y, Dai J, Yu X, Li L, El-Serag HB, Kanwal F. Effect of diabetes medications and glycemic control on risk of hepatocellular cancer in patients with nonalcoholic fatty liver disease. Hepatology 2022;75:1420-1428.

37.   Lee TY, Wu JC, Yu SH, Lin JT, Wu MS, Wu CY. The occurrence of hepatocellular carcinoma in different risk stratifications of clinically noncirrhotic nonalcoholic fatty liver disease. Int J Cancer 2017;141:1307-1314.

38.   Mao X, Zhang X, Kam L, Chien N, Lai R, Cheung KS, Yuen MF, et al. Synergistic association of sodium-glucose cotransporter-2 inhibitor and metformin on liver and non-liver complications in patients with type 2 diabetes mellitus and metabolic dysfunction-associated steatotic liver disease. Gut 2024.

39.   Roudier E, Mistafa O, Stenius U. Statins induce mammalian target of rapamycin (mTOR)-mediated inhibition of Akt signaling and sensitize p53-deficient cells to cytostatic drugs. Mol Cancer Ther 2006;5:2706-2715.

40.   Relja B, Meder F, Wang M, Blaheta R, Henrich D, Marzi I, Lehnert M. Simvastatin modulates the adhesion and growth of hepatocellular carcinoma cells via decrease of integrin expression and ROCK. Int J Oncol 2011;38:879-885.

41.   Sutter AP, Maaser K, Höpfner M, Huether A, Schuppan D, Scherübl H. Cell cycle arrest and apoptosis induction in hepatocellular carcinoma cells by HMG-CoA reductase inhibitors. Synergistic antiproliferative action with ligands of the peripheral benzodiazepine receptor. J Hepatol 2005;43:808-816.

42.   Kah J, Wüstenberg A, Keller AD, Sirma H, Montalbano R, Ocker M, Volz T, et al. Selective induction of apoptosis by HMG-CoA reductase inhibitors in hepatoma cells and dependence on p53 expression. Oncol Rep 2012;28:1077-1083.

43. German MN, Lutz MK, Pickhardt PJ, Bruce RJ, Said A. Statin Use is Protective Against Hepatocellular Carcinoma in Patients With Nonalcoholic Fatty Liver Disease: A Case-control Study. J Clin Gastroenterol 2020;54:733-740.

44. Pinyopornpanish K, Al-Yaman W, Butler RS, Carey W, McCullough A, Romero-Marrero C. Chemopreventive Effect of Statin on Hepatocellular Carcinoma in Patients With Nonalcoholic Steatohepatitis Cirrhosis. Am J Gastroenterol 2021;116:2258-2269.

45. Vilar-Gomez E, Calzadilla-Bertot L, Wai-Sun Wong V, Castellanos M, Aller-de la Fuente R, Metwally M, Eslam M, et al. Fibrosis Severity as a Determinant of Cause-Specific Mortality in Patients With Advanced Nonalcoholic Fatty Liver Disease: A Multi-National Cohort Study. Gastroenterology 2018;155:443-457.e417.

46. Marra F, Svegliati-Baroni G. Lipotoxicity and the gut-liver axis in NASH pathogenesis. J Hepatol 2018;68:280-295.

47. Fuchs A, Samovski D, Smith GI, Cifarelli V, Farabi SS, Yoshino J, Pietka T, et al. Associations Among Adipose Tissue Immunology, Inflammation, Exosomes and Insulin Sensitivity in People With Obesity and Nonalcoholic Fatty Liver Disease. Gastroenterology 2021;161:968-981.e912.

48. Friedman SL, Neuschwander-Tetri BA, Rinella M, Sanyal AJ. Mechanisms of NAFLD development and therapeutic strategies. Nat Med 2018;24:908-922.

49. Daugherity EK, Balmus G, Al Saei A, Moore ES, Abi Abdallah D, Rogers AB, Weiss RS, et al. The DNA damage checkpoint protein ATM promotes hepatocellular apoptosis and fibrosis in a mouse model of non-alcoholic fatty liver disease. Cell Cycle 2012;11:1918-1928.

50. Ng SWK, Rouhani FJ, Brunner SF, Brzozowska N, Aitken SJ, Yang M, Abascal F, et al. Convergent somatic mutations in metabolism genes in chronic liver disease. Nature 2021;598:473-478.

51. Zhu C, Kim K, Wang X, Bartolome A, Salomao M, Dongiovanni P, Meroni M, et al. Hepatocyte Notch activation induces liver fibrosis in nonalcoholic steatohepatitis. Sci Transl Med 2018;10.

52. Zhu C, Tabas I, Schwabe RF, Pajvani UB. Maladaptive regeneration - the reawakening of developmental pathways in NASH and fibrosis. Nat Rev Gastroenterol Hepatol 2021;18:131-142.

53.   Lee KC, Wu PS, Lin HC. Pathogenesis and treatment of non-alcoholic steatohepatitis and its fibrosis. Clin Mol Hepatol 2023;29:77-98.

54.   Sanyal AJ, Van Natta ML, Clark J, Neuschwander-Tetri BA, Diehl A, Dasarathy S, Loomba R, et al. Prospective Study of Outcomes in Adults with Nonalcoholic Fatty Liver Disease. N Engl J Med 2021;385:1559-1569.

55.   Ramachandran P, Dobie R, Wilson-Kanamori JR, Dora EF, Henderson BEP, Luu NT, Portman JR, et al. Resolving the fibrotic niche of human liver cirrhosis at single-cell level. Nature 2019;575:512-518.

56.   Zhang DY, Friedman SL. Fibrosis-dependent mechanisms of hepatocarcinogenesis. Hepatology 2012;56:769-775.

57.   Lei H, Reinke P, Volk HD, Lv Y, Wu R. Mechanisms of Immune Tolerance in Liver Transplantation-Crosstalk Between Alloreactive T Cells and Liver Cells With Therapeutic Prospects. Front Immunol 2019;10:2667.

58.   Kitade M, Yoshiji H, Kojima H, Ikenaka Y, Noguchi R, Kaji K, Yoshii J, et al. Neovascularization and oxidative stress in the progression of non-alcoholic steatohepatitis. Mol Med Rep 2008;1:543-548.

59.   Coulon S, Legry V, Heindryckx F, Van Steenkiste C, Casteleyn C, Olievier K, Libbrecht L, et al. Role of vascular endothelial growth factor in the pathophysiology of nonalcoholic steatohepatitis in two rodent models. Hepatology 2013;57:1793-1805.

60.   Kwak M, Mehaffey JH, Hawkins RB, Hsu A, Schirmer B, Hallowell PT. Bariatric surgery is associated with reduction in non-alcoholic steatohepatitis and hepatocellular carcinoma: A propensity matched analysis. Am J Surg 2020;219:504-507.

61.   Yan M, Man S, Ma L, Guo L, Huang L, Gao W. Immunological mechanisms in steatotic liver diseases: An overview and clinical perspectives. Clin Mol Hepatol 2024.

62.   Sutti S, Albano E. Adaptive immunity: an emerging player in the progression of NAFLD. Nat Rev Gastroenterol Hepatol 2020;17:81-92.

63.   Wolf MJ, Adili A, Piotrowitz K, Abdullah Z, Boege Y, Stemmer K, Ringelhan M, et al. Metabolic activation of intrahepatic CD8+ T cells and NKT cells causes nonalcoholic steatohepatitis and liver cancer via cross-talk with hepatocytes. Cancer Cell 2014;26:549-564.

64.   Heinrich B, Brown ZJ, Diggs LP, Vormehr M, Ma C, Subramanyam V, Rosato U, et al. Steatohepatitis Impairs T-cell-Directed Immunotherapies Against Liver Tumors in Mice. Gastroenterology 2021;160:331-345.e336.

65.   Ma C, Kesarwala AH, Eggert T, Medina-Echeverz J, Kleiner DE, Jin P, Stroncek DF, et al. NAFLD causes selective CD4(+) T lymphocyte loss and promotes hepatocarcinogenesis. Nature 2016;531:253-257.

66.   Deczkowska A, David E, Ramadori P, Pfister D, Safran M, Li B, Giladi A, et al. XCR1(+) type 1 conventional dendritic cells drive liver pathology in non-alcoholic steatohepatitis. Nat Med 2021;27:1043-1054.

67.   Ou R, Liu J, Lv M, Wang J, Wang J, Zhu L, Zhao L, et al. Neutrophil depletion improves diet-induced non-alcoholic fatty liver disease in mice. Endocrine 2017;57:72-82.

68.   Leslie J, Mackey JBG, Jamieson T, Ramon-Gil E, Drake TM, Fercoq F, Clark W, et al. CXCR2 inhibition enables NASH-HCC immunotherapy. Gut 2022;71:2093-2106.

69.   McVey JC, Green BL, Ruf B, McCallen JD, Wabitsch S, Subramanyam V, Diggs LP, et al. NAFLD indirectly impairs antigen-specific CD8(+) T cell immunity against liver cancer in mice. iScience 2022;25:103847.

70.   Wabitsch S, McCallen JD, Kamenyeva O, Ruf B, McVey JC, Kabat J, Walz JS, et al. Metformin treatment rescues CD8(+) T-cell response to immune checkpoint inhibitor therapy in mice with NAFLD. J Hepatol 2022;77:748-760.

71.   Dudek M, Pfister D, Donakonda S, Filpe P, Schneider A, Laschinger M, Hartmann D, et al. Auto-aggressive CXCR6(+) CD8 T cells cause liver immune pathology in NASH. Nature 2021;592:444-449.

72.   Pfister D, Núñez NG, Pinyol R, Govaere O, Pinter M, Szydlowska M, Gupta R, et al. NASH limits anti-tumour surveillance in immunotherapy-treated HCC. Nature 2021;592:450-456.

73.   Ma C, Han M, Heinrich B, Fu Q, Zhang Q, Sandhu M, Agdashian D, et al. Gut microbiome-mediated bile acid metabolism regulates liver cancer via NKT cells. Science 2018;360.

74.   Jadhav K, Cohen TS. Can You Trust Your Gut? Implicating a Disrupted Intestinal Microbiome in the Progression of NAFLD/NASH. Front Endocrinol (Lausanne) 2020;11:592157.

75.   Parthasarathy G, Revelo X, Malhi H. Pathogenesis of Nonalcoholic Steatohepatitis: An Overview. Hepatol Commun 2020;4:478-492.

76.   Mouries J, Brescia P, Silvestri A, Spadoni I, Sorribas M, Wiest R, Mileti E, et al. Microbiota-driven gut vascular barrier disruption is a prerequisite for non-alcoholic steatohepatitis development. J Hepatol 2019;71:1216-1228.

77.   Boursier J, Mueller O, Barret M, Machado M, Fizanne L, Araujo-Perez F, Guy CD, et al. The severity of nonalcoholic fatty liver disease is associated with gut dysbiosis and shift in the metabolic function of the gut microbiota. Hepatology 2016;63:764-775.

78.   Madrid AM, Hurtado C, Venegas M, Cumsille F, Defilippi C. Long-Term treatment with cisapride and antibiotics in liver cirrhosis: effect on small intestinal motility, bacterial overgrowth, and liver function. Am J Gastroenterol 2001;96:1251-1255.

79.   Liu YL, Patman GL, Leathart JB, Piguet AC, Burt AD, Dufour JF, Day CP, et al. Carriage of the PNPLA3 rs738409 C >G polymorphism confers an increased risk of non-alcoholic fatty liver disease associated hepatocellular carcinoma. J Hepatol 2014;61:75-81.

80.   Liu YL, Reeves HL, Burt AD, Tiniakos D, McPherson S, Leathart JB, Allison ME, et al. TM6SF2 rs58542926 influences hepatic fibrosis progression in patients with non-alcoholic fatty liver disease. Nat Commun 2014;5:4309.

81.   Donati B, Dongiovanni P, Romeo S, Meroni M, McCain M, Miele L, Petta S, et al. MBOAT7 rs641738 variant and hepatocellular carcinoma in non-cirrhotic individuals. Sci Rep 2017;7:4492.

82.   Kawaguchi T, Shima T, Mizuno M, Mitsumoto Y, Umemura A, Kanbara Y, Tanaka S, et al. Risk estimation model for nonalcoholic fatty liver disease in the Japanese using multiple genetic markers. PLoS One 2018;13:e0185490.

83.   Bianco C, Jamialahmadi O, Pelusi S, Baselli G, Dongiovanni P, Zanoni I, Santoro L, et al. Non-invasive stratification of hepatocellular carcinoma risk in non-alcoholic fatty liver using polygenic risk scores. J Hepatol 2021;74:775-782.

84.	Hoshida Y, Nijman SM, Kobayashi M, Chan JA, Brunet JP, Chiang DY, Villanueva A, et al. Integrative transcriptome analysis reveals common molecular subclasses of human hepatocellular carcinoma. Cancer Res 2009;69:7385-7392.

85.	Pinyol R, Torrecilla S, Wang H, Montironi C, Piqué-Gili M, Torres-Martin M, Wei-Qiang L, et al. Molecular characterisation of hepatocellular carcinoma in patients with non-alcoholic steatohepatitis. J Hepatol 2021;75:865-878.

86.	Jühling F, Hamdane N, Crouchet E, Li S, El Saghire H, Mukherji A, Fujiwara N, et al. Targeting clinical epigenetic reprogramming for chemoprevention of metabolic and viral hepatocellular carcinoma. Gut 2021;70:157-169.

87.	Singh S, Singh PP, Singh AG, Murad MH, Sanchez W. Statins are associated with a reduced risk of hepatocellular cancer: a systematic review and meta-analysis. Gastroenterology 2013;144:323-332.

88.	Simon TG, Duberg AS, Aleman S, Chung RT, Chan AT, Ludvigsson JF. Association of Aspirin with Hepatocellular Carcinoma and Liver-Related Mortality. N Engl J Med 2020;382:1018-1028.

89.	EASL-EASD-EASO Clinical Practice Guidelines on the management of metabolic dysfunction-associated steatotic liver disease (MASLD). J Hepatol 2024;81:492-542.

90.	Kang SH, Lee HW, Yoo JJ, Cho Y, Kim SU, Lee TH, Jang BK, et al. KASL clinical practice guidelines: Management of nonalcoholic fatty liver disease. Clin Mol Hepatol 2021;27:363-401.

91.	Rinella ME, Tacke F, Sanyal AJ, Anstee QM. Report on the AASLD/EASL Joint Workshop on Clinical Trial Endpoints in NAFLD. Hepatology 2019;70:1424-1436.

92.	Baumeister SE, Schlesinger S, Aleksandrova K, Jochem C, Jenab M, Gunter MJ, Overvad K, et al. Association between physical activity and risk of hepatobiliary cancers: A multinational cohort study. J Hepatol 2019;70:885-892.

93.	Promrat K, Kleiner DE, Niemeier HM, Jackvony E, Kearns M, Wands JR, Fava JL, et al. Randomized controlled trial testing the effects of weight loss on nonalcoholic steatohepatitis. Hepatology 2010;51:121-129.

94.	Lange NF, Radu P, Dufour JF. Prevention of NAFLD-associated HCC: Role of lifestyle and chemoprevention. J Hepatol 2021;75:1217-1227.

95.   Aby E, Phan J, Truong E, Grotts J, Saab S. Inadequate Hepatocellular Carcinoma Screening in Patients With Nonalcoholic Steatohepatitis Cirrhosis. J Clin Gastroenterol 2019;53:142-146.

96.   Wolf E, Rich NE, Marrero JA, Parikh ND, Singal AG. Use of Hepatocellular Carcinoma Surveillance in Patients With Cirrhosis: A Systematic Review and Meta-Analysis. Hepatology 2021;73:713-725.

97.   Tan DJH, Ng CH, Lin SY, Pan XH, Tay P, Lim WH, Teng M, et al. Clinical characteristics, surveillance, treatment allocation, and outcomes of non-alcoholic fatty liver disease-related hepatocellular carcinoma: a systematic review and meta-analysis. Lancet Oncol 2022;23:521-530.

98.   Chong N, Schoenberger H, Yekkaluri S, Fetzer DT, Rich NE, Yokoo T, Gopal P, et al. Association between ultrasound quality and test performance for HCC surveillance in patients with cirrhosis: a retrospective cohort study. Aliment Pharmacol Ther 2022;55:683-690.

99.   Schoenberger H, Chong N, Fetzer DT, Rich NE, Yokoo T, Khatri G, Olivares J, et al. Dynamic Changes in Ultrasound Quality for Hepatocellular Carcinoma Screening in Patients With Cirrhosis. Clin Gastroenterol Hepatol 2022;20:1561-1569.e1564.

100.   Tzartzeva K, Obi J, Rich NE, Parikh ND, Marrero JA, Yopp A, Waljee AK, et al. Surveillance Imaging and Alpha Fetoprotein for Early Detection of Hepatocellular Carcinoma in Patients With Cirrhosis: A Meta-analysis. Gastroenterology 2018;154:1706-1718.e1701.

101.   Yu JH, Lee HA, Kim SU. Non-invasive imaging biomarkers for liver fibrosis in nonalcoholic fatty liver disease: current and future. Clin Mol Hepatol 2023;29:S136-s149.

102.   Koh YX, Tan HJ, Liew YX, Syn N, Teo JY, Lee SY, Goh BKP, et al. Liver Resection for Nonalcoholic Fatty Liver Disease-Associated Hepatocellular Carcinoma. J Am Coll Surg 2019;229:467-478.e461.

103.   Foerster F, Gairing SJ, Müller L, Galle PR. NAFLD-driven HCC: Safety and efficacy of current and emerging treatment options. J Hepatol 2022;76:446-457.

104.   Petrelli F, Cortellini A, Indini A, Tomasello G, Ghidini M, Nigro O, Salati M, et al. Association of Obesity With Survival Outcomes in Patients With Cancer: A Systematic Review and Meta-analysis. JAMA Netw Open 2021;4:e213520.

105.   Wang YG, Wang P, Wang B, Fu ZJ, Zhao WJ, Yan SL. Diabetes mellitus and poorer prognosis in hepatocellular carcinoma: a systematic review and meta-analysis. PLoS One 2014;9:e95485.

106.   Su CW, Chau GY, Hung HH, Yeh YC, Lei HJ, Hsia CY, Lai CR, et al. Impact of Steatosis on Prognosis of Patients with Early-Stage Hepatocellular Carcinoma After Hepatic Resection. Ann Surg Oncol 2015;22:2253-2261.

107.   Molinari M, Kaltenmeier C, Samra PB, Liu H, Wessel C, Lou Klem M, Dharmayan S, et al. Hepatic Resection for Hepatocellular Carcinoma in Nonalcoholic Fatty Liver Disease: A Systematic Review and Meta-Analysis of 7226 Patients. Ann Surg Open 2021;2:e065.

108.   Chin KM, Prieto M, Cheong CK, Di Martino M, Ielpo B, Goh BKP, Koh YX. Outcomes after curative therapy for hepatocellular carcinoma in patients with non-alcoholic fatty liver disease: a meta-analysis and review of current literature. HPB (Oxford) 2021;23:1164-1174.

109.   Wong RJ, Chou C, Bonham CA, Concepcion W, Esquivel CO, Ahmed A. Improved survival outcomes in patients with non-alcoholic steatohepatitis and alcoholic liver disease following liver transplantation: an analysis of 2002-2012 United Network for Organ Sharing data. Clin Transplant 2014;28:713-721.

110.   Haldar D, Kern B, Hodson J, Armstrong MJ, Adam R, Berlakovich G, Fritz J, et al. Outcomes of liver transplantation for non-alcoholic steatohepatitis: A European Liver Transplant Registry study. J Hepatol 2019;71:313-322.

111.   Kern B, Feurstein B, Fritz J, Fabritius C, Sucher R, Graziadei I, Bale R, et al. High incidence of hepatocellular carcinoma and postoperative complications in patients with nonalcoholic steatohepatitis as a primary indication for deceased liver transplantation. Eur J Gastroenterol Hepatol 2019;31:205-210.

112.   Wong CR, Njei B, Nguyen MH, Nguyen A, Lim JK. Survival after treatment with curative intent for hepatocellular carcinoma among patients with vs without non-alcoholic fatty liver disease. Aliment Pharmacol Ther 2017;46:1061-1069.

113.   Young S, Sanghvi T, Rubin N, Hall D, Roller L, Charaf Y, Golzarian J. Transarterial Chemoembolization of Hepatocellular Carcinoma: Propensity Score Matching Study Comparing Survival and

Complications in Patients with Nonalcoholic Steatohepatitis Versus Other Causes Cirrhosis. Cardiovasc Intervent Radiol 2020;43:65-75.

114.   Schotten C, Bechmann LP, Manka P, Theysohn J, Dechêne A, El Fouly A, Barbato F, et al. NAFLD-Associated Comorbidities in Advanced Stage HCC Do Not Alter the Safety and Efficacy of Yttrium-90 Radioembolization. Liver Cancer 2019;8:491-504.

115.   Haber PK, Puigvehí M, Castet F, Lourdusamy V, Montal R, Tabrizian P, Buckstein M, et al. Evidence-Based Management of Hepatocellular Carcinoma: Systematic Review and Meta-analysis of Randomized Controlled Trials (2002-2020). Gastroenterology 2021;161:879-898.

116.   Kelley RK, Rimassa L, Cheng AL, Kaseb A, Qin S, Zhu AX, Chan SL, et al. Cabozantinib plus atezolizumab versus sorafenib for advanced hepatocellular carcinoma (COSMIC-312): a multicentre, open-label, randomised, phase 3 trial. Lancet Oncol 2022;23:995-1008.

117.   Song YG, Yoo JJ, Kim SG, Kim YS. Complications of immunotherapy in advanced hepatocellular carcinoma. J Liver Cancer 2024;24:9-16.

118.   Finn RS, Qin S, Ikeda M, Galle PR, Ducreux M, Kim TY, Kudo M, et al. Atezolizumab plus Bevacizumab in Unresectable Hepatocellular Carcinoma. N Engl J Med 2020;382:1894-1905.

119.   Yau T, Kaseb A, Cheng AL, Qin S, Zhu AX, Chan SL, Melkadze T, et al. Cabozantinib plus atezolizumab versus sorafenib for advanced hepatocellular carcinoma (COSMIC-312): final results of a randomised phase 3 study. Lancet Gastroenterol Hepatol 2024;9:310-322.

120.   Abou-Alfa GK, Lau G, Kudo M, Chan SL, Kelley RK, Furuse J, Sukeepaisarnjaroen W, et al. Tremelimumab plus Durvalumab in Unresectable Hepatocellular Carcinoma. NEJM Evid 2022;1:EVIDoa2100070.

121.   Yau T, Park JW, Finn RS, Cheng AL, Mathurin P, Edeline J, Kudo M, et al. Nivolumab versus sorafenib in advanced hepatocellular carcinoma (CheckMate 459): a randomised, multicentre, open-label, phase 3 trial. Lancet Oncol 2022;23:77-90.

122.   Qin S, Kudo M, Meyer T, Bai Y, Guo Y, Meng Z, Satoh T, et al. Tislelizumab vs Sorafenib as First-Line Treatment for Unresectable Hepatocellular Carcinoma: A Phase 3 Randomized Clinical Trial. JAMA Oncol 2023;9:1651-1659.

123.   Llovet JM, Kudo M, Merle P, Meyer T, Qin S, Ikeda M, Xu R, et al. Lenvatinib plus pembrolizumab versus lenvatinib plus placebo for advanced hepatocellular carcinoma (LEAP-002): a randomised, double-blind, phase 3 trial. Lancet Oncol 2023;24:1399-1410.

124.   Qin S, Chan SL, Gu S, Bai Y, Ren Z, Lin X, Chen Z, et al. Camrelizumab plus rivoceranib versus sorafenib as first-line therapy for unresectable hepatocellular carcinoma (CARES-310): a randomised, open-label, international phase 3 study. Lancet 2023;402:1133-1146.

125.   Ren Z, Xu J, Bai Y, Xu A, Cang S, Du C, Li Q, et al. Sintilimab plus a bevacizumab biosimilar (IBI305) versus sorafenib in unresectable hepatocellular carcinoma (ORIENT-32): a randomised, open-label, phase 2-3 study. Lancet Oncol 2021;22:977-990.

126.   Finn RS, Ryoo BY, Merle P, Kudo M, Bouattour M, Lim HY, Breder V, et al. Pembrolizumab As Second-Line Therapy in Patients With Advanced Hepatocellular Carcinoma in KEYNOTE-240: A Randomized, Double-Blind, Phase III Trial. J Clin Oncol 2020;38:193-202.

127.   Qin S, Chen Z, Fang W, Ren Z, Xu R, Ryoo BY, Meng Z, et al. Pembrolizumab Versus Placebo as Second-Line Therapy in Patients From Asia With Advanced Hepatocellular Carcinoma: A Randomized, Double-Blind, Phase III Trial. J Clin Oncol 2023;41:1434-1443.

128.   Castet F, Willoughby CE, Haber PK, Llovet JM. Atezolizumab plus Bevacizumab: A Novel Breakthrough in Hepatocellular Carcinoma. Clin Cancer Res 2021;27:1827-1829.

129.   Bruix J, Raoul JL, Sherman M, Mazzaferro V, Bolondi L, Craxi A, Galle PR, et al. Efficacy and safety of sorafenib in patients with advanced hepatocellular carcinoma: subanalyses of a phase III trial. J Hepatol 2012;57:821-829.

130.   Cheng AL, Kang YK, Chen Z, Tsao CJ, Qin S, Kim JS, Luo R, et al. Efficacy and safety of sorafenib in patients in the Asia-Pacific region with advanced hepatocellular carcinoma: a phase III randomised, double-blind, placebo-controlled trial. Lancet Oncol 2009;10:25-34.

131.   Cheng AL, Guan Z, Chen Z, Tsao CJ, Qin S, Kim JS, Yang TS, et al. Efficacy and safety of sorafenib in patients with advanced hepatocellular carcinoma according to baseline status: subset analyses of the phase III Sorafenib Asia-Pacific trial. Eur J Cancer 2012;48:1452-1465.

132.   Kudo M, Finn RS, Qin S, Han KH, Ikeda K, Piscaglia F, Baron A, et al. Lenvatinib versus sorafenib in first-line treatment of patients with unresectable hepatocellular carcinoma: a randomised phase 3 non-inferiority trial. Lancet 2018;391:1163-1173.

133.   Abou-Alfa GK, Meyer T, Cheng AL, El-Khoueiry AB, Rimassa L, Ryoo BY, Cicin I, et al. Cabozantinib in Patients with Advanced and Progressing Hepatocellular Carcinoma. N Engl J Med 2018;379:54-63.

134.   Bruix J, Qin S, Merle P, Granito A, Huang YH, Bodoky G, Pracht M, et al. Regorafenib for patients with hepatocellular carcinoma who progressed on sorafenib treatment (RESORCE): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet 2017;389:56-66.

135.   Zhu AX, Kang YK, Yen CJ, Finn RS, Galle PR, Llovet JM, Assenat E, et al. Ramucirumab after sorafenib in patients with advanced hepatocellular carcinoma and increased α-fetoprotein concentrations (REACH-2): a randomised, double-blind, placebo-controlled, phase 3 trial. Lancet Oncol 2019;20:282-296.

Table 1. Summary of key phase III randomized trials evaluating the efficacy and safety of systemic therapies according to the etiology of hepatocellular carcinoma

Trial	Treatment arms	Subgroup based on etiology (n, %)	Overall survival HR (95% CI)
Immunotherapy: first line			
IMbrave150[118]	Atezolizumab plus bevacizumab vs. sorafenib	Overall (501)	0.58 (0.42–0.79)
		HBV (240, 48%)	0.51 (0.32–0.81)
		HCV (108, 22%)	0.43 (0.22–0.87)
		Non-viral (153, 31%)	0.91 (0.52–1.60)
COSMIC-312[119]	Atezolizumab plus cabozantinib vs. sorafenib	Overall (649)	0.90 (0.69–1.18)
		HBV (190, 29%)	0.53 (0.33–0.87)

		HCV (202, 31%)	1.10 (0.72–1.68)
		Non-viral (257, 40%)	1.18 (0.78–1.79)
HIMALAYA[120]	Tremelimumab plus durvalumab vs. sorafenib	Overall (782)	0.78 (0.65–0.93)
		HBV (241, 31%)	0.64 (0.48–0.86)
		HCV (214, 27%)	1.06 (0.76–1.49)
		Non-viral (327, 42%)	0.74 (0.57–0.95)
	Durvalumab vs. sorafenib	Overall (778)	0.86 (0.73–1.03)
		HBV (238, 31%)	0.78 (0.58–1.04)
		HCV (211, 27%)	1.05 (0.75–1.48)
		Non-viral (329, 42%)	0.82 (0.64–1.05)
CheckMate 459[121]	Nivolumab vs. sorafenib	Overall (743)	0.85 (0.72–1.02)
		HBV (233, 31%)	0.77 (0.56–1.05)
		HCV (173, 23%)	0.71 (0.49–1.01)
		Non-viral (336, 45%)	0.95 (0.74–1.22)
RATIONALE-301[122]	Tislelizumab vs. sorafenib	Overall (674)	0.85 (0.71–1.02)
		HBV (427, 63%)	0.91 (0.73–1.14)
		HCV (85, 13%)	0.64 (0.38–1.08)
		Non-viral (162, 24%)	0.78 (0.55–1.12)
LEAP-002[123]	Pembrolizumab plus lenvatinib vs. lenvatinib	Overall (794)	0.84 (0.71–1.00)
		HBV (385, 47%)	0.75 (0.58–0.97)
		HCV (181, 22%)	0.86 (0.60–1.24)
		Alcohol (251, 31%)	0.84 (0.67–1.05)
CARES-310[124]	Camrelizumab plus rivoceranib vs. sorafenib	Overall (543)	0.62 (0.49–0.80)
		HBV (405, 75%)	0.66 (0.50–0.87)
		HCV (51, 9%)	0.45 (0.18–1.16)

		Non-viral (87, 16%)	0.71 (0.37–1.36)
ORIENT-32[125]	Sintilimab plus IBI305 vs. sorafenib	Overall (571)	0.57 (0.43–0.75)
		HBV (538, 94%)	0.58 (0.43–0.76)
		Non-HBV (33, 6%)	0.80 (0.22–2.87)
Immunotherapy: second line			
KEYNOTE-240[126]	Pembrolizumab vs. placebo	Overall (413)	0.78 (0.61–1.00)
		HBV (101, 24%)	0.57 (0.35–0.94)
		HCV (64, 15%)	0.96 (0.48–1.92)
		Non-viral (248, 60%)	0.88 (0.64–1.20)
KEYNOTE-394[127]	Pembrolizumab vs. placebo	Overall (453)	0.79 (0.63–0.99)
		HBV (360, 79%)	0.78 (0.61–0.99)
		Non-HBV (93, 21%)	0.87 (0.53–1.44)
Tyrosine kinase inhibitors: first line			
SHARP[128, 129]	Sorafenib vs. placebo	Overall (439)	0.69 (0.55–0.87)
		HBV (111, 18%)	0.76 (0.38–1.50)
		HCV (169, 28%)	0.50 (0.32–0.77)
		Alcohol (159, 26%)	0.76 (0.50–1.16)
Asia–Pacific[130, 131]	Sorafenib vs. placebo	Overall (226)	0.68 (0.50–0.93)
		HBV (165, 73%)	0.74 (0.51–1.06)
		Non-HBV (61, 27%)	0.57 (0.29–1.13)
REFLECT[132]	Lenvatinib vs. sorafenib	Overall (753)	0.92 (0.79–1.06)
		HBV (479, 50%)	0.83 (0.68–1.02)
		HCV (217, 23%)	0.91 (0.66–1.26)
		Alcohol (57, 6%)	1.03 (0.47–2.28)
Tyrosine kinase inhibitors: second line			

CELESTIAL[133]	Cabozantinib vs. placebo	Overall (707)	0.76 (0.63–0.92)
		HBV (267, 38%)	0.69 (0.51–0.94)
		HCV (168, 24%)	1.11 (0.72–1.71)
		Non-viral (272, 38%)	0.72 (0.54–0.96)
RESORCE[134]	Regorafenib vs. placebo	Overall (573)	0.63 (0.50–0.79)
		HBV (216, 38%)	0.58 (0.41–0.82)
		HCV (119, 21%)	0.79 (0.49–1.26)
		Alcohol (145, 25%)	0.92 (0.61–1.38)
REACH-2[135]	Ramucirumab vs. placebo	Overall (292)	0.71 (0.53–0.95)
		HBV (107, 37%)	0.84 (0.52–1.35)
		HCV (76, 26%)	0.76 (0.44–1.33)
		Other (109, 37%)	0.63 (0.38–1.06)

HCC, hepatocellular carcinoma; HR, hazard ratio; HBV, hepatitis B virus; HCV, hepatitis C virus
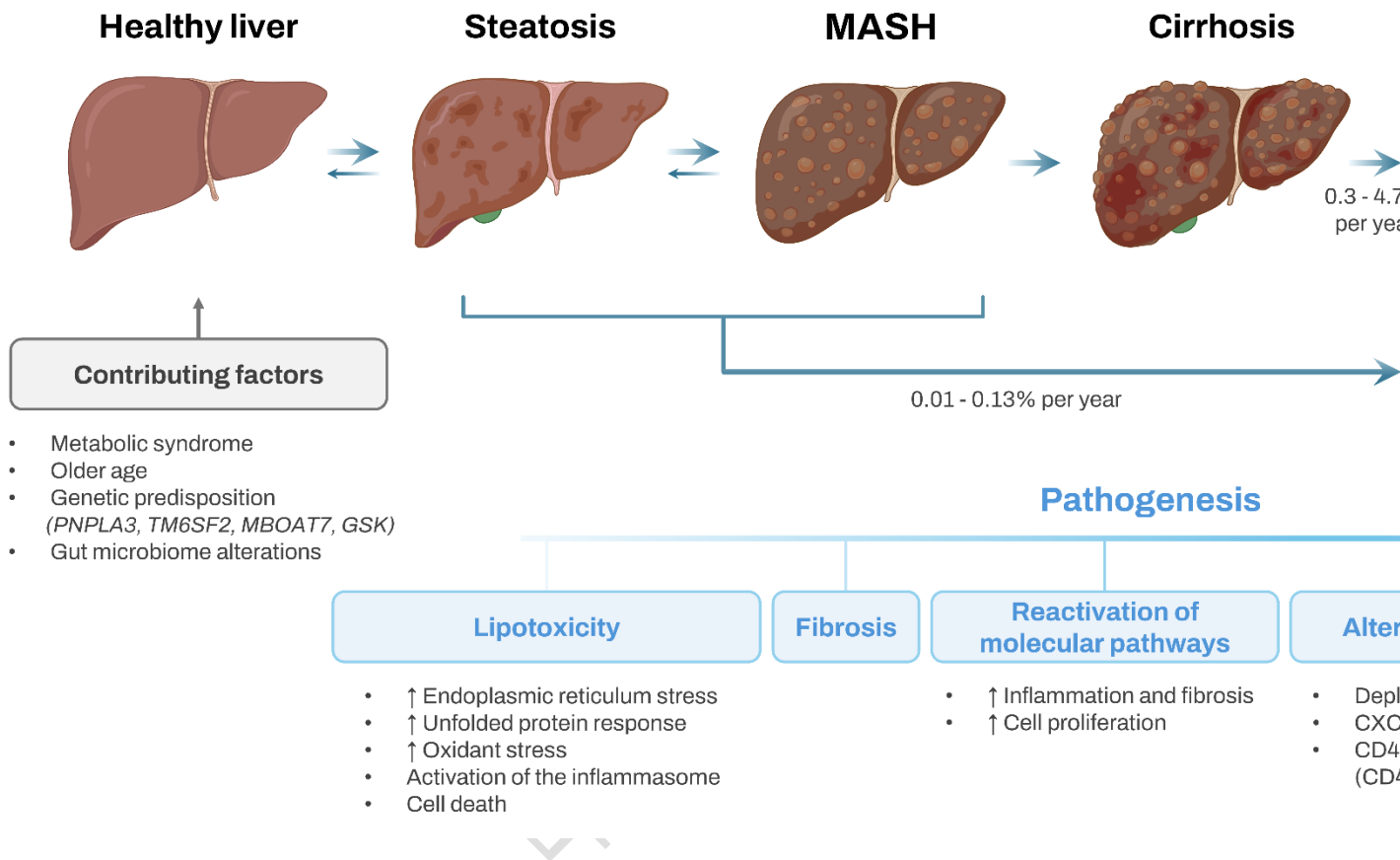
Fig. 1. Pathogenesis and progression of MASH-HCC (Drawn by the author).

MASH: metabolic dysfunction-associated steatohepatitis; HCC: hepatocellular carcinoma; T_H: T helper